



Role of Matrix Metalloproteinases in Angiogenesis and Cancer

Saray Quintero-Fabián^{1†}, Rodrigo Arreola^{2†}, Enrique Becerril-Villanueva³, Julio César Torres-Romero⁴, Victor Arana-Argáez⁵, Julio Lara-Riegos⁴, Mario Alberto Ramírez-Camacho⁶ and María Elizabeth Alvarez-Sánchez^{7*}

¹ Multidisciplinary Research Laboratory, Military School of Graduate of Health, Mexico City, Mexico, ² Psychiatric Genetics Department, National Institute of Psychiatry "Ramón de la Fuente", Clinical Research Branch, Mexico City, Mexico, ³ Psychoimmunology Laboratory, National Institute of Psychiatry "Ramón de la Fuente", Mexico City, Mexico, ⁴ Biochemistry and Molecular Genetics Laboratory, Facultad de Química de la Universidad Autónoma de Yucatán, Mérida, Mexico, ⁵ Pharmacology Laboratory, Facultad de Química de la Universidad Autónoma de Yucatán, Mérida, Mexico, ⁶ Centro de Información de Medicamentos, Facultad de Química de la Universidad Autónoma de Yucatán, Mérida, Mexico, ⁷ Genomic Sciences Graduate Program, Universidad Autónoma de la Ciudad de México, Mexico City, Mexico

OPEN ACCESS

Edited by:

Erika Ruiz-García,
National Institute of Cancerology
(INCan), Mexico

Reviewed by:

Linda C. Meade-Tollin,
University of Arizona, United States
Ronca Roberto,
University of Brescia, Italy

*Correspondence:

María Elizabeth Alvarez-Sánchez
maria.alvarez@uacm.edu.mx

[†]These authors have contributed
equally to this work

Specialty section:

This article was submitted to
Molecular and Cellular Oncology,
a section of the journal
Frontiers in Oncology

Received: 24 June 2019

Accepted: 20 November 2019

Published: 06 December 2019

Citation:

Quintero-Fabián S, Arreola R,
Becerril-Villanueva E,
Torres-Romero JC, Arana-Argáez V,
Lara-Riegos J, Ramírez-Camacho MA
and Alvarez-Sánchez ME (2019) Role
of Matrix Metalloproteinases in
Angiogenesis and Cancer.
Front. Oncol. 9:1370.
doi: 10.3389/fonc.2019.01370

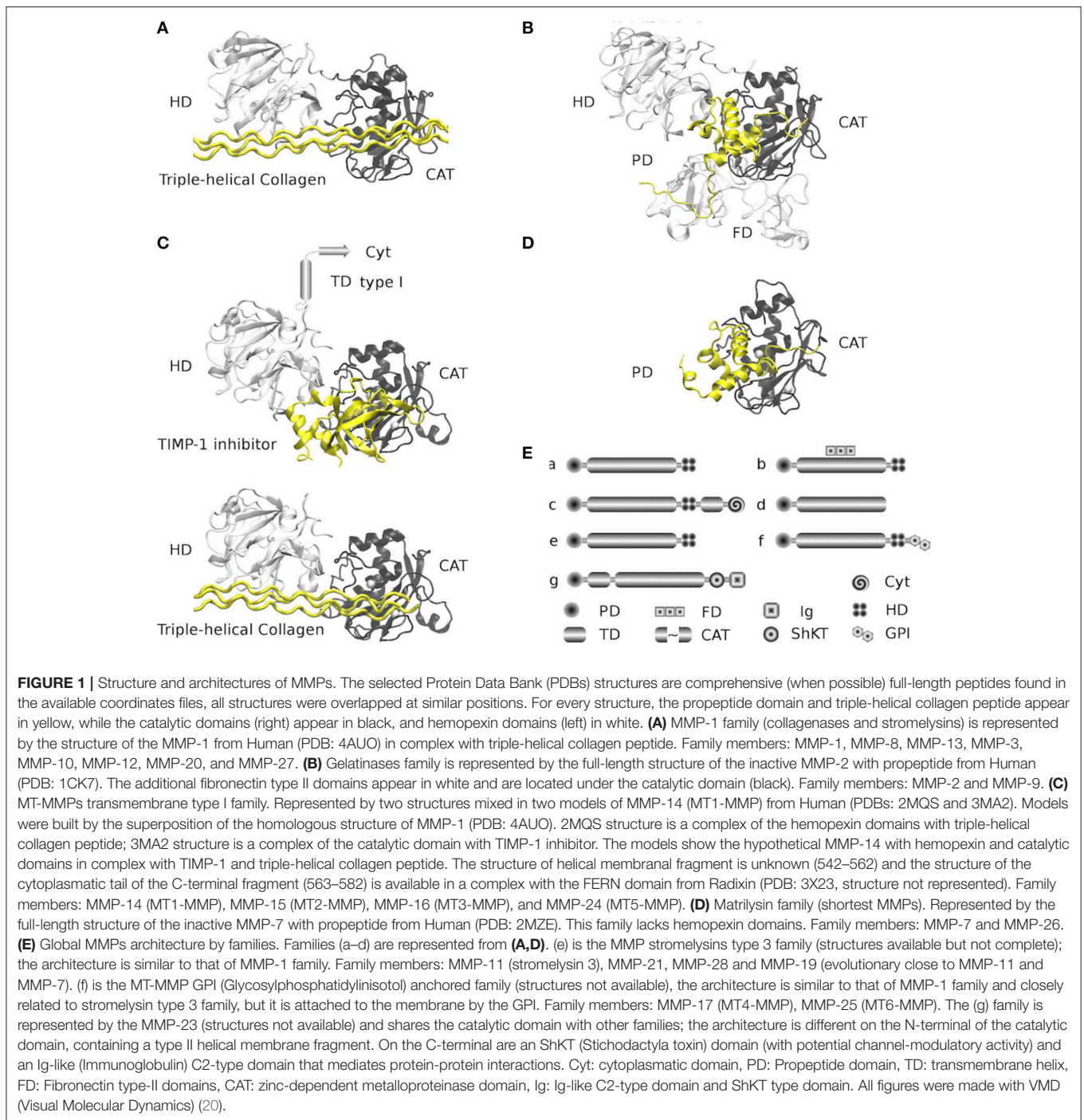
During angiogenesis, new vessels emerge from existing endothelial lined vessels to promote the degradation of the vascular basement membrane and remodel the extracellular matrix (ECM), followed by endothelial cell migration, and proliferation and the new generation of matrix components. Matrix metalloproteinases (MMPs) participate in the disruption, tumor neovascularization, and subsequent metastasis while tissue inhibitors of metalloproteinases (TIMPs) downregulate the activity of these MMPs. Then, the angiogenic response can be directly or indirectly mediated by MMPs through the modulation of the balance between pro- and anti-angiogenic factors. This review analyzes recent knowledge on MMPs and their participation in angiogenesis.

Keywords: angiogenesis and cancer, immune system, metalloproteinases, MMP, MT-MMP

INTRODUCTION

Epithelial-Mesenchymal Transition (EMT) in Metastasis and Migration

Currently, cancer research is focused on understanding the functional mechanisms underlying cell transformation and tumor progression that can be used to develop new markers and therapies (1). Cancer metastasis, the final step of tumor progression and the leading cause of cancer morbidity and mortality, involves the spread of cancer cells from the primary tumor to nearby tissues and distant organs; it is mediated by complex molecular changes of in cell cycle regulation (2, 3). The molecular changes that regulate the cell morphology and functions of epithelial cells, that is epithelial-mesenchymal transition (EMT), include the destruction of intercellular relationships and cell-matrix adhesive characteristics, extracellular matrix (ECM) breakdown, and cleavage of basement membrane components by matrix metalloproteinase (MMP) activity modulation. For example, when epithelial cells lose their polarity through EMT, cell-cell tight junctions and adhesive connections are lost, resulting in infiltration and an enhanced migration ability of these cells (4, 5). Therefore, EMT enables malignant cells to become motile and invasive, which constitutes a fundamental requisite for cancer metastasis (6).

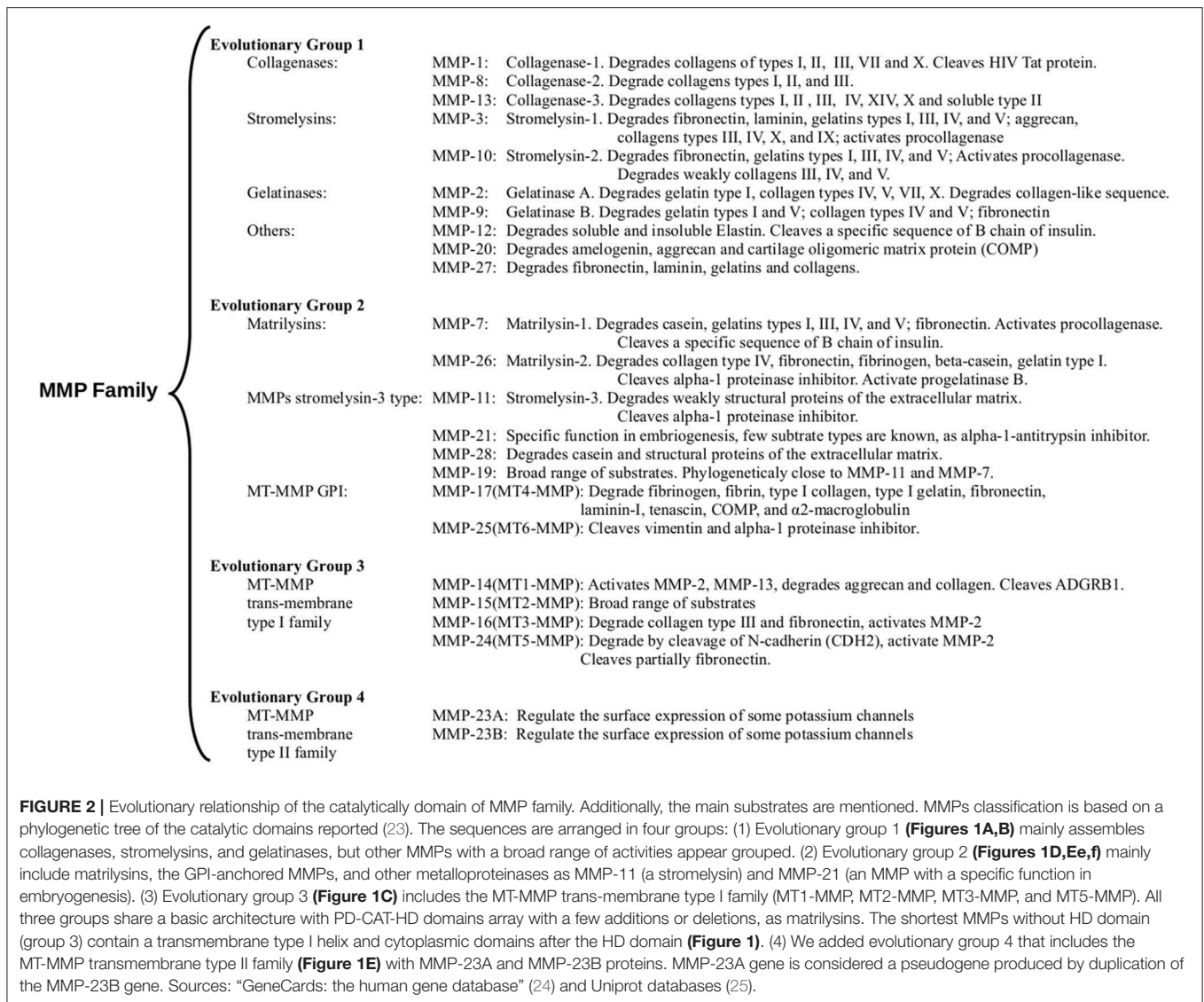


a cytokine VEGF homolog that stimulates angiogenesis and various types of cells, such as myeloids and stromals cancers, in addition to activating tumor cells, while their inhibition improves cancer treatment (33).

Collagenases (MMP-1, –8, and –13) are proteins associated with angiogenesis, and their loss leads to the irreversible rupture of the matrix. Type IV collagen participates in cell endothelial migration in the interstitial stromal spaces. It is known that the

tissue inhibitors of metalloproteinases (TIMP-1, TIMP-2, TIMP-3, and TIMP-4) regulate them, playing a key role in angiogenesis regulation by inhibiting neovascularization (34).

In adults, angiogenesis is initiated only under inflammation or hypoxic conditions (35). In the early proliferative stage, vascular repair must predominate to control bleeding by vasoconstriction and coagulation. During menstruation, the endometrium is expelled if the ovule is not fertilized. Women who suffer from



endometriosis show aggressive angiogenesis in the peritoneal cavity (36).

On the other hand, several studies have established the importance of transmembrane receptors and ligands participating in cell differentiation. Their role in endothelial sprouting during angiogenesis has recently been studied. ECs express several Notch receptors (such Notch1 and Notch4), as well as the Notch1 protein and Notch ligand delta-like 4 (DLL4), which are important signals for vascular development (37). In most of the healthy population, resting ECs showed long half-lives through VEGF activation, Notch signaling, and angiopoietin-1 (ANG-1) and fibroblast growth factors (FGFs) expression (33). Recent knowledge concerning the complexity of angiogenesis indubitably shows the role of the participants in this event and allows for finding applications in anti-angiogenic therapy.

As previously mentioned, angiogenesis is a normal development and part of the healing process; however, it is

key to tumor branching and arborization under pathological conditions such as cancer. The formation of new vascular networks promotes the growth, maintenance, and spread of cancer (38). During angiogenesis in cancer, alterations have been described at the level of lymphangiogenesis and vasculogenesis, both processes are highly involved in the propagation of cancer cells and an unfavorable prognosis (39).

The accelerated growth of the tumor leads to hypoxic tumor microenvironment, interstitial hypertension, and acidosis. To reverse these adverse physicochemical changes, VEGF-C and VEGF-D are synthesized by the activation of VEGFR-3/2, triggering a rise in diameter and density of the peritumoral lymphatic vessels, favoring the propagation of tumor cells toward sentinel lymph nodes (40, 41). It has been shown that the inhibition of these factors by the use of antibodies decreases lymphogenesis and metastasis in nearby ganglia (42–44). Then, angiogenesis maintains a constant and permanent supply of nutrients for cancer cells that leads to tumor growth. This

aberrant revascularization begins after the loss of regulation of inhibitory factors (e.g., thrombospondin-1) and angiogenic promoters (VEGF) (45, 46). Hypoxia-inducible factor (HIF) is one of the first growth factors to initiate the abnormal process of vascular growth and responds to the low oxygen tension in the tumor mass. Subsequently, a wave of growth factors such as EGF, basic and acidic FGF, estrogen, prostaglandin E1 and E2, IL-8, TGF, TNF, neuropilins, and VEGF promotes the formation of a vascular network that ensures the exchange of oxygen and nutrients with the tumor (5, 31, 47, 48). This vascularization process is regulated primarily by VEGF-A/VEGF-1,2 and DLL4 signaling. The activation of ECs also triggers a branching process toward the central region of the tumor (49, 50). This new supply and drainage network that supports the tumor allows the latter to maintain a favorable microenvironment for its growth and dissemination. At present, the tumor niche is considered an independent organ able to maintain itself (51). Additionally, integrin receptors are overexpressed in tumor ECs and play a key role connecting the cell cytoskeleton to the extracellular matrix protein ligands such as arginine-glycine-aspartic acid (RGD). This binding interaction between integrin and protein ligands is an important mechanism during the angiogenesis of tumor endothelial cells (52).

MMPs in Cancer Angiogenesis

It is well known that MMPs have been implicated in angiogenesis regulation as well as in the anomalous relationship between cancer and the related processes of angiogenesis, vasculogenesis and lymphangiogenesis. MMPs also have a role in the immune system action in cancer development and progression (Tables 1, 2). The pro- and anti-angiogenic effects of MMPs participate in crucial steps as the ability to degrade ECM or cleave several substrates. Specifically, MMP-2 and MMP-9 give rise to the modulation of the dynamic remodeling of ECM (editing aggrecan, collagens, elastin, fibronectin, laminins, and glycosaminoglycans, and latent signaling proteins), activating and deactivating by proteolytic cleavages releasing biological activities that induce cellular regulation (108, 109). MMP activation can be induced by several angiogenic factors, such as VEGF, basic fibroblast growth factors (bFGF), TGF- α and - β , and angiogenin. Specifically, MMP-1 activity promotes the expression of the vascular endothelial growth factor receptor 2 (VEGFR2) and EC proliferation, stimulating serine/threonine-protein kinase MARK2 (PAR-1) and activating the transcription factor NF- κ B, suggesting the existence of a mechanism by which MMP-1 stimulates vascular remodeling and angiogenesis (110). Similarly, MMP-7 modulates the VEGF pathway in human umbilical vein endothelial cells (HUVECs), degrading soluble VEGFR-1 and in turn promoting angiogenesis (111). TNF- α , IL-8 and other factors with a known pro-angiogenic capacity, stimulate the production of MMP-2, -8, and -9 in ECs and regulate the angiogenesis process (63, 112).

Angiogenesis studies using MMP-8 and MMP-2 knock-out mice, show an *in vitro* reduction of cell proliferation and neocapillary network growth, as well as a decrease in HUVEC migration and poor *in vivo* angiogenesis. Interestingly, ischemia-induced neovascularization is also affected by a reduction

in ECs and invasive, proliferative, or mobilizing activities of endothelial progenitor cells (EPCs) derived from bone marrow (113, 114). Additionally, it is known that ECs secrete MMP-2 and -9-containing vesicles stimulated by VEGF and FGF-2 and thus regulate the proteolytic activity critical for the angiogenesis-related invasive and morphogenic processes (115). Furthermore, MMP-9 also generates the angiogenic and tumoral repressor, tumstatin by proteolysis of the non collagenous domain (NC1) from the collagen alpha-3(IV) chain. The anti-angiogenic properties of tumstatin inhibit EC proliferation and induce apoptosis by interacting with alphaVbeta3 integrin (116).

Among the most studied MMPs participating in angiogenesis is the MMP-14 (MT1-MMP). It significantly contributes to angiogenesis regulation by cleaving ECM molecules as a matrix-degrading enzyme (Figure 3). This MMP also acts as a key effector in the production of pro-angiogenic factors such as VEGF. In addition, MT1-MMP interacts with cell surface molecules, such as CD44 and sphingosine 1-phosphate receptor 1 (S1P₁), to induce EC migration, and plays a critical role in the proteolytic degradation of anti-angiogenic factors as decorin. Furthermore, evidence shows that MT1-MMP is able to degrade pro-TGF- β and endoglin (TGF- β receptor), suggesting a pivotal role in vessel maturation and angiogenesis, respectively (117) (Figure 3A). In addition, MT1-MMP appears to be an essential molecule that determines ECM adhesion and human endothelial cell tube formation through the modulation of MMP-2 expression (Figure 3B). This suggests an important role in regulating angiogenesis-related functions in human ECs (118).

Soluble MMPs in Cancer Angiogenesis

Soluble MMP expression and its effects on cancer stabilization/proliferation are intimately linked via vascular angiogenesis mechanisms that are now well recognized. In this regard, MMP-1 expression has been reported to contribute to the progression of Head and neck squamous cell carcinomas (HNSCC) and the suggest metastatic phenotype of human breast and colorectal cancers, among others (119–121). Interestingly, MMP-1/protease-activated receptor-1 (PAR1) signaling axis has been implicated in tumor angiogenesis and intravasation of carcinoma cells by inducing vascular permeability (122), as well as, hypoxia-regulated MMP-1 expression in metastatic bladder cancer cells, which could be associated to a reactive oxygen species (ROS)-related regulation of the spheroid metastatic phenotype and cell spread (123). The increased expression of MMP-1 in human chondrosarcoma is an important prognostic factor and its function in the spread of tumor cells has been evaluated by silencing assays in which cancer metastasis is impaired but local tumor growth and angiogenesis are enhanced (124). These findings strongly support a role for MMP-1 in the diverse proliferative outcomes of human cancer through angiogenic processes.

Many studies have been published describing the relationship between MMP-2 expression and tumor angiogenesis. One of the earliest reports indicates that IL-8, an angiogenic factor, induces MMP-2 expression and activity in melanoma cells, enhancing their invasion (125). A relationship between MMP-2 expression and stromal support, angiogenesis, invasiveness, and tumor

TABLE 1 | Immune system proteins associated to MMPs in angiogenesis and cancer.

Protein	Intervention/process	Tumor type/cell lines	MMP	References
IL-1 α , IL-3, VEGF, GCSF and GM-CSF	Secretion of proteins (IL-1 α , IL-3, VEGF, GCSF, GM-CSF) under Hypoxia stress increase efficiency for induction of angiogenesis	Human A431 squamous carcinoma cells Human A549 non-small lung cells, and H1299 NSCL lung cells	Hypoxia increase secretion: MMP-13. Hypoxia increase secretion: MMP-3, MMP-9 and MMP-13	(53)
IL-1 β TNF- α	IL-1 β as inductor shows a slight dose-dependent increasing secretion of MMP-2. TNF- α as inductor shows a slight dose-dependent increasing secretion of MMP-9. A curious fact, retinoic acid strongly inhibited MMP-2 secretion	Human Glioblastoma T-98G cell line	MMP-2 and MMP-9	(54)
IL-1 β	IL-1 β induced MMP-2 and MMP-9 expression and activities mediated NK-kB activation, whereas melatonin suppresses it	Human Gastric adenocarcinoma MGC803 cell line and Human Gastric cancer SGC-7901 cell line	MMP-2 and MMP-9	(19)
IL-1 β	IL-1 β /p38/AP-1(c-fos)/MMP-2 and MMP-9 pathway play an important role in metastasis in gastric cancer	Human Gastric cancer cell lines MKN45 and AGS	MMP-2 and MMP-9 increase gene expression and protein expression in response to IL-1 β treatment	(55)
IL-1 β	The STAT3 signaling is present in myeloid cells in human cancer angiogenesis and it is required for the cellular migration. The activity of STAT3 in tumor-associated myeloid cells participate in the elevated gene transcription of VEGF, bFGF, IL-1 β MMP-9, CCL2 and CXL2	Murine Tumor-infiltrating myeloid cells	MMP-9 is elevated by the STAT3 activity	(23)
IL-5	IL-5 increased migration and MMP-9 expression via activation of transcription factors NF- κ B and AP-1, and induced activation of ERK1/2 and Jak-Stat signaling in both cells. IL-5R α , inhibition, suppressed migration, ERK1/2, NF- κ B, AP-1 activation and MMP-9 expression. MMP-2 expression remains without changes	Human Bladder carcinoma cell lines: 5637, T24 and HT1376	MMP-2 and MMP-9	(24, 25)
IL-22 (IL-10 family member) and IL-22R1	Promotes gastric cancer cell invasion through STAT3 and ERK signaling in MKN28 Promotes gastric cancer cell invasion activating AKT signaling in SGC-7901	Human Gastric cancer cell lines MKN28 and SGC-7901	IL-22 upregulate the gene expression of MMP-7 and MMP-13 in MKN28 IL-22 upregulate the gene expression of MMP-9 in SGC-7901	(56, 57)
IL-10	IL-10-stimulated macrophages polarized to M2 phenotype (low IL-12, IL-6 expression and IL-10 high expression) significantly increased AGS and RKO cells Invasion radio. Conditioned medium from IL-10-stimulated macrophages (M2) induced in AGS cell motility, migration and mediated angiogenesis	Human Diffuse gastric carcinoma cell line: AGS Colon carcinoma cell line: RKO	MMP-2 and MMP-9 elevated expression and activities on AGS cells with conditioned medium from IL-10-stimulated macrophages (M2)	(58)
IL-8 (CXCL8) IL-1 β	Breast cancer cells secreting high levels of RANTES, CCL2 and G-CSF showing a potential capability to recruit monocytes and to instruct them to secrete high levels of IL-1 β and IL-8, and MMP-1, MMP-2 and MMP-10	Patient samples diagnosed with ductal carcinoma. Monocytic Cell lines THP-1, U937, and Human Breast cancer cell lines: T47D (HTB-133), MCF-7 and MDA-MB-231	MMP-1, MMP-2 and MMP-10	(59)
IL-8 (CXCL8)	Co-cultured ovarian cancer stem-like cells with macrophages (derived from THP-1 cells) polarized to M2 phenotype increased IL-10, VEGF, MMP-9 and IL-8 secretion, and CD163 and STAT3 expression. THP-1 cell conditioned medium plus IL-8 induced stemness in SKOV3 cells involving IL-8/STAT3 signaling.	Human SKOV3-derived ovarian cancer stem-like cells.	MMP-9	(60)
IL-8 (CXCL8)	Recruited B cells mediated IL-8/androgen receptor and MMP signals in bladder cancer could enhance invasion and metastasis	Bladder tumor specimens were collected from 24 patients Human Bladder cancer cell lines TCCSUP, T24 and J82, and the Ramos B cell line	MMP-1 and MMP-9	(61)

(Continued)

TABLE 1 | Continued

Protein	Intervention/process	Tumor type/cell lines	MMP	References
IL-8 (CXCL8)	<i>Porphyromonas gingivalis</i> on chronic periodontitis promoted the invasive ability of carcinoma cells by up-regulation of IL-8, MMP-1 and MMP-2. Other MMPs are up-regulated too like MMP-7, MMP-9 and MMP-10	Human Oral squamous cell carcinoma cells SCC-25, OSC-20 and SAS cells	MMP-1, MMP-2, MMP-7, MMP-9 and MMP-10 are up-regulated after 72 hours of <i>P. gingivalis</i> infection	(62)
IL-8 (CXCL8)	IL-8 directly enhances endothelial cell survival, proliferation, MMP production and modulate angiogenesis	Human Umbilical Vein Endothelial Cell and dermal microvascular endothelial cells	MMP-2 and MMP-9 mRNA expression was increased in cells treated with 10 and 100 ng/ml IL-8. The Culture supernatant showed high level of both active MMPs	(63)
IL-8 (CXCL8) IL-6	IL-8, IL-9, MMP-2 and MMP-9 secreted by Falconi Anemia Cells are expressed under the control of NF- κ B/TNF- α signaling pathways. These secretory factors are effective on promoting proliferation, migration, invasion of surrounding tumor cells	Falconi Anemia Cells (EUFA274, EUFA274Rev, EUFA450, EUFA450RevR, FANCD2 and FANCD2 corrected), MDA-MB-231 cells, PC3 cells, MCF7 cells, MCF10A cells	MMP-2 MMP-7 and MMP-9 are overexpressed	(64)
IL-8 (CXCL8) IL-1 β , VEGF	Self-conditioned medium collected from A549 cells was treated with neutralizing antibodies against IL-1 β , IL-8, and VEGF and used in A459 cells. The inhibition of motility and invasion in A549 cells were observed, the effect was higher in IL-8 and VEGF neutralizing medium	A549 (human lung adenocarcinoma), MCF-7 (breast adenocarcinoma) and HT-29 (colon carcinoma)	MMP-2 activity was detected in Self-conditioned medium collected from A549 cells	(65)
IL-8, VEGF, angiogenin, and NKG2D	Lung tumor-associated NK cells (TANKs) of peripheral blood and tumor-infiltrating NK cells (TINKs) induced functional angiogenesis-associated behaviors of endothelial cells <i>in vitro</i> . TANKs release TIMP-1, TIMP-2, and MMP-9, proteins involved in tissue remodeling and invasion process. They also increase the expression the angiogenin, VEGF and CXCL8, depending on tumor location	Human Lung tumor-associated NK cells (TANKs) of peripheral blood and tumor-infiltrating NK cells (TINKs) of patients with colorectal cancer	MMPs down-regulate the activator NKG2D, a surface marker for NK cell activation, in TANKs. Which is correlated with increased release of MMP-9, TIMP-1 and TIMP-2.	(66)
IL-8, IL-6, IL-1a, IL-1RA, GM-CSF, CCL5 (RANTES), TNF- α , VEGFA	Different lines cells from similar tumors show a varied secreted immunological biomarker profiles. Although almost every cells lines express the eight cytokines, apparently the metastatic stage, cellular origin, the site and the genome differences plus, an uncertain passage number of the cell lines, cause different profiles: SCC25 express mostly VEGFA and CCL5; SCC19 express mostly VEGFA, IL-6 and IL-8; SCC92 mostly express TNF- α IL-6, IL-1 α and IL-8; SCC99 express mostly IL-8	Human Head and neck squamous cell carcinoma lines: SCC4, SCC15, SCC25, SCC84 and SCC92 are from the oral cavity; while SCC19 and SCC99 are from the oropharynx	MMP-1, MMP-7 and MMP-9 are higher expressed on SCC25. Others cell lines express different MMP profiles: SCC99 mostly express MMP-1 and MMP-9; SCC15, SCC19 and SCC84 mostly express MMP-7 and MMP-9; SCC4 and SCC92 mostly express MMP-9	(67)
IL-8	IL-8 and MMP-9 are co-expressed on MCF-7 cell line induced by TPA (a carcinogen). Orientin downregulates signal PKC α /ERK and blocks the nuclear translocation of AP-1 and STAT3 causing an attenuation of IL-8 and MMP-9 induced by TPA treatment, but only affected the migration and invasion of ER-positive MCF-7 cells	Human Breast cancer cell line MCF-7 estrogen receptor positive	MMP-9	(68)
IL-8, IL-6	MMP expression is regulated by cancer cell density via the signaling of IL-6 and IL-8. The synergistic signaling of IL-6 and IL-8 regulates the production of MMPs through the JAK/STAT signaling pathway	Human Fibrosarcoma HT1080 cells and breast carcinoma MDA-MB-231 cells	HT1080 in high cell density not only expresses MMP-1, MMP-2 and MMP-3 mainly but also MMP-11 and MMP-14 MDA-MB-231 in high cell density not only expresses MMP-14 mainly, but also MMP-7, MMP-1 and MMP-2	(69)

(Continued)

TABLE 1 | Continued

Protein	Intervention/process	Tumor type/cell lines	MMP	References
IL-6	In macrophages, the homeo-domain protein Six1 overexpression was able to induce IL-6 up-regulation and increase activity of STAT3 in Hepatocellular carcinoma cells. Macrophages Six1 upregulate IL-6 and MMP-9 and can stimulate cancer cell invasion by elevating MMP-9 expression	Human Leukemic monocyte cell line: THP-1; Human hepatoma cell line: A59T; and hepatocellular carcinoma cell line: HepG2 Two paraffin-embedded Hepatocellular carcinoma tissue arrays	MMP-9	(70)
IL-6	IL-6 regulates MMP expression via proximal GAS-like STAT binding elements (SBEs). IL-6 lead the formation of a complex STAT1/AP-1	Patient colon tumor tissue Human Colorectal carcinoma cell lines HT29, SW480, LISP-1, LIM1215, HCT116, and LS174T Hepatocyte-derived HepG2 cell line	MMP-1 and MMP-3	(71)
IL-6	IL-6/ NOS2 inflammatory signals regulate MMP-9 and MMP-2 dependent metastatic activity	Nasopharyngeal carcinoma from patients	MMP-9 and MMP-2	(72)
IL-6	IL-6 secreted by astrocytes induce upregulation of MMP-14 increasing migration and invasion of Glioma cell lines	Human Glioma cell lines U251 and A172 Astrocytes	MMP-14 (MT1-MMP)	(73)
IL-11	IL-11 promoting chronic gastric inflammation and associated tumorigenesis mediated by excessive activation of STAT3 and STAT1	Gastric tumor gp130 Y757F/Y757F mice model	Upregulate the gene expression of MMP-13	(74)
IL-11	Under hypoxia conditions all cell lines upregulate gene expression and protein production of IL-11 Recombinant IL-11 treatment increased the migration and invasion under normoxia and neutralizing antibodies and siRNA suppressed the migration and invasion. Recombinant IL-11 increased STAT3 phosphorylation and expression of MMP-2, MMP-3 and MMP-9	Human Breast cancer cell line: MDA-MB-231; colorectal carcinoma cell line: HCT116; non-small lung carcinoma: H1299; malignant melanoma cell line: A375 and hepatocellular carcinoma cell line: HepG2	Upregulate the gene expression of MMP-2, MMP-3 and MMP-9	(75)
IL-12	IL-12 treatment inhibited lung tumor growth, resulting in the long-term survival of lung cancer-bearing mice. Further examination revealed that IL-12 rapidly activated NK cells to secrete IFN- γ , resulting in the inhibition of tumor angiogenesis and MMP-9 transcript level decreased MMP-9/TIM modulation in the tumor microenvironment after 14 days of IL-12 therapy produce reversible antitumoral effects	Murine breast cancer HTH-K (syngeneic breast carcinoma), injected in C57BL/6 mice to generate an orthotopic lung cancer model	L-12 prevented blood vessel regrowth and inhibit MMP-9	(76, 77)
IL-17	In breast tumors was observed the presence of IL-17 strongly positive cells within the scattered tumor-associated inflammatory infiltrate. IL-17 addition to breast cancer cell lines promoted significant invasiveness	Human Archival paraffin-embedded sections of 19 primary invasive breast tumors (15 Grade III and four Grade II). Human Breast cancer cell lines: MDA-MB231 and MDAMB435 cell lines	Selective antagonists for MMP-2/MMP-9 or MMP-3 suppressed the stimulatory effect of IL-17 on breast cancer invasion. However, IL-17 does not affect secretion of these MMPs	(78, 79)
IL-17	High salt synergizes with sub-effective IL-17 to induce breast cancer cell proliferation mediated activation of SIK3 (a G0/G1-phase inducer) by mTOR complex. SIK3 induce expression of CXCR4 through MMP-9 activation	Human Breast cancer cells lines: MCF7, MDA-MB-231, BT20, AU565	MMP-9	(80)
IL-17	MMP-7 mediates IL-17's function in promoting prostate carcinogenesis through induction of EMT, indicating IL-17-MMP-7-EMT axis as potential targets for developing new strategies in the prevention and treatment of prostate cancer	Murine Prostate cancer cell lines (LNCaP, C4-2B and PC-3). PB-Cre4 mice	MMP-7	(81)

(Continued)

TABLE 1 | Continued

Protein	Intervention/process	Tumor type/cell lines	MMP	References
IL-17B, IL-17RB	IL-17B dose dependently promoted the invasion, growth and migration of thyroid cancer cells. IL-17RB induced ERK1/2 activation pathway and increased MMP-9 expression Thyroid cancer cells express IL-17RA, IL-17RB and IL-17RC	16 paired Human thyroid cancer tissues Thyroid epithelial cell line HTori-3 and the thyroid cancer cell lines TPC-1, SW1736 and FTC-133	MMP-9	(82)
IL-17	IL-17A treatment promotes OE19 cell migration and invasion, upregulates MMP-2 and MMP-9 expression, increase ROS production, I κ B- α phosphorylation and NF- κ B nuclear translocation. IL-17 cause these effects through ROS/NF- κ B/MMP-2/9 signaling pathway	Human Esophagus adenocarcinoma Cell line OE19	MMP-2 and MMP-9	(83)
IL-18, IL-10 and TNF- α	IL-18 and IL-10 synergistically act to amplify OPN and thrombin production, which in turn augments M2 macrophage polarization. M2 Macrophages and endothelial direct cell-cell interaction resulting in excessive angiogenesis	Mouse leukemic monocyte Mphi cell line RAW264.7 and Mouse endothelial cell line b.End5	Stimulation of RAW264.7 cells with TNF- α increases MMP-2 and MMP-9 gene expression IL-18 and/or IL-10 had no impact in the gene expression of MMP-2, MMP-3, MMP-7 and MMP-9	(84)
IL-32 α	IL-32 stimulation in MG-63 cells shown, dose-dependently promoted the invasion and motility of osteosarcoma cells and induced the activation of AKT in a time-dependent manner. IL-32 stimulation increased the expression and secretion of MMP-13	Human MG-63 osteosarcoma cell line	MMP-13	(85)
IL-33	IL-33 increases the abilities of proliferation, migration and invasion of melanoma cells and Vasculogenic mimicry tube formation through ST2. IL-33 induces the production of MMP-2/9 via ERK1/2 phosphorylation	Human Melanoma of patients	MMP-2 and MMP-9	(86)
IL-33	IL-33 significantly promoted cell invasion and migration and induced the expression of MMP-2 and MMP-9 via ST2 and AKT pathway	Human Lung cancer cell lines: A549 and NCI-H1299	MMP-2 and MMP-9	(87)
IL-33, IL1RL1 (IL-1-R4)	IL-33 expression in the tumor epithelium of adenomas and carcinomas and expression of the IL-33 receptor, its receptor IL1RL1 in the stroma of adenoma and both the stroma and epithelium of human colorectal cancer IL-33 signaling promotes tumor growth and associated angiogenesis	Human colorectal cancer and mouse model of intestinal tumorigenesis	MMP-1 and MMP-3	(88, 89)
IL-35	IL-35 can induce N2 neutrophil polarization (protumor phenotype) by increasing G-CSF and IL-6 production, and promote Neutrophil infiltration into tumor microenvironment. IL-35 stimulated macrophages to secrete proinflammatory cytokines IL-1 β , IL-6 and up-regulate the expression of MMP-9, suggesting antitumoral activity	Murine H22 hepatocarcinoma cell Line and B16F0 melanoma cells	MMP-9	(90)
IL-35	Significantly lower expression of IL-35 was also observed in Hepatocellular carcinoma patients. IL-35 over-expression in HepG2 cells significantly upregulated HLA-ABC and CD95, reduced activities of MMP-2 and MMP-9, and decreased cell migration, invasion and colony formation capacities	Hepatocellular carcinoma from 75 patients and Human Hepatocellular carcinoma cell line HepG2	MMP-2 and MMP-9	(91)
IL-37	Transfected cells A549 overexpressing IL-37 cause low gene expression of MMP-9, PCNA, Ki-67, Cyclin D1 and CDK4, but elevated	Murine Lung adenocarcinoma line cells A549. Xenograft mouse models.	MMP-9	(92)

(Continued)

TABLE 1 | Continued

Protein	Intervention/process	Tumor type/cell lines	MMP	References
	expression of caspase-3 and caspase-9. IL-37 inhibits the proliferation, migration and invasion of human lung adenocarcinoma A549 cells as well as the chemotaxis of Treg cells and promotes apoptosis of A549 cells			
CXCR4	Lymph node metastatic Hepatocarcinome Hca-F exosomes (contain elevated CXCR4) promote migration and invasion in HcaP cells elevating the secretion of MMP-9, MMP-2 and VEGF-C	Murine hepatocarcinoma cell lines Hca-F and Hca-P	MMP-2 and MMP-9	(93)
TNF- α	Tumor necrosis factor- α (TNF- α) induce a dose-dependent increase in MMP-9 activity HT1376 cells, through ERK1/2 and P38 MAP kinase activity and activation of the transcription factors NF- κ B, AP-1 and SP-1	Human Bladder carcinoma cell line, HT1376	MMP-9 induced by TNF- α through the NF- κ B, AP-1 and Sp-1 cis-elements of the gene promoter mediated regulation ERK1/2 and p38 MAP kinase	(94)
TNF- α	TNF- α secretion from cancer cell line increased expression of MMP-2 and MMP-9 and increased TNF- α production. A TNF- α /TNF-R1/NF- κ B system signaling pathway generated a highly metastatic cancer cells. TNF- α -triggered NF- κ B activation to upregulation of active MMP released from the cancer cells	Human Oral squamous cell carcinoma SAS cell line. Metastatic cervical lymph nodes and metastatic lung cell lines induced by a SAS injected in the tongue of mouse	MMP-2 and MMP-9	(95)
TNF- α	Aberrant TNF- α signaling promotes cancer cell motility, invasiveness, and enhances cancer metastasis mediated NF- κ B signaling. TNF- α -induced expression and stabilization of C/EBP β depends on p38MAPK activation, but not on NF- κ B activity. C/EBP β and its downstream MMP-1 and MMP-3 are required for TNF- α -induced cancer cell migration. TNF- α activates multiple signaling pathways, including NF- κ B and C/EBP β to promote cancer cell migration. TNF- α treatment significantly increased the number of migrated MDA-MB-231 and MDA-MB-435 cells in a dose-dependent manner	Human Breast cancer lines cells: MDA-MB-231 and MDA-MB-435	MMP-1 and MMP-3 mediate TNF- α -induced cell migration downstream of C/EBP β	(96)
TNF- α	AMB cells stimulation with TNF- α increased IL-6 and MMP-9 mRNA expressions, via NF- κ B activation. Furthermore, TGF- β and IFN- γ increased TNF- α -mediated expressions of MMP-9 and IL-6 mRNA, while those responses were suppressed by NF- κ B inhibitor	Ameloblastoma cells (AMB) cultures from patients were immortalized using hTERT vector	MMP-9	(97)

growth was demonstrated using an MMP-2-specific inhibitor in a mouse model of bladder cancer (126). Furthermore, an elevated expression of MMP-2 was correlated with VEGF expression in gastric cancer (127) which suggests that this MMP plays a critical role in the progression of cancer through ECM degradation, tumor neovascularization and metastasis.

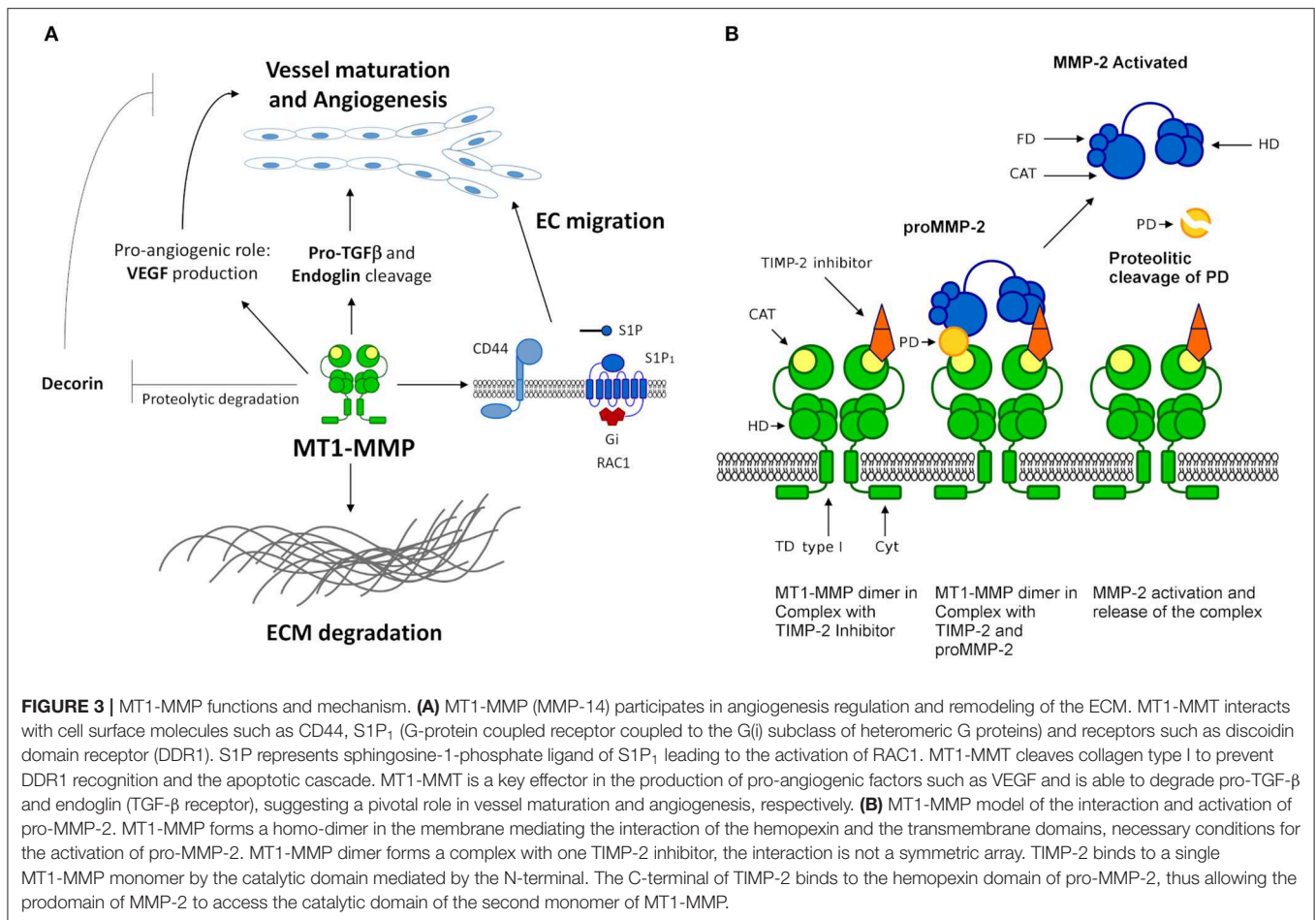
On the other hand, MMP-9 promotes endothelial cell migration and triggers the angiogenic switch by releasing VEGF during carcinogenesis (128). Decreased expression of VEGF and MMP-9 in medulloblastoma cells that overexpress osteonectin, also referred to as Secreted Protein Acidic and Rich in Cysteine (SPARC), leads to decreased angiogenesis and tumor growth, indicating the pro-angiogenic role of MMP-9 in cancer tissues (129). In contrast, the direct proteolytic cleavage of osteopontin (OPN) by MMP-9 contributes to cancer metastasis, most likely

associated with angiogenesis via the regulation of VEGF and angiostatin secretion (130, 131). This model suggests that cancer growth is accompanied by increased vascular permeability, due in part to the expression of MMP-9, leading to the regulation of angiogenic factors, and eventually, neovascularization in cancer tissue.

Studies have revealed that, both MMP-2 and MMP-9 can degrade type IV collagen and are frequently elevated in human cancer. Additionally, a cooperative effect of MMP-2 and MMP-9 was demonstrated in an *in vivo* experimental model establishing the angiogenic phenotype and invasiveness of tumor keratinocytes (132). The mechanism whereby MMP-2 and MMP-9 activity induces cancer angiogenesis involves the cleavage of latent TGF- β in a CD44-dependent manner, which can promote tumor growth and invasion (133). Together, these results confirm

TABLE 2 | Proteins associated with MMPs in angiogenesis on cancer.

Protein	Intervention/process	Tumor type	MMP	References
L1 adhesion molecule /CD171	Constitutive cleavage of L1 proceeds in exosomes mediated by a disintegrin and MMP10, under apoptotic conditions multiple MMP are involved	Human ovarian carcinoma cells OVMz	ADAM10	(98)
PIGF	Knockdown of PIGF in spheroid body cells reduced <i>in vitro</i> tumorigenicity and stemness properties (self-renewal ability, colony forming, migration and MMPs activities) and decreased ability to differentiation and angiogenesis	Human Spheroid cells from gastric adenocarcinoma MKN-45 and GS cells lines	MMP-2 and MMP-9 activities	(99)
VEGFR2 blockade	Brain tumor vessels: Vascular stabilization by increases pericyte coverage, up-regulation of angiopoietin-1 and collagenase IV activity provides and oxygenated environment through the degrades pathologically thick basement membrane by MMPs activation	Human Orthotopic glioblastoma obtained by xenografts on mouse of U87 gliomas tumors	MMPs–2 and MMP-9	(100)
VEGF	In colorectal liver metastasis, the high expression of stroma-derived MMP-12 and VEGF correlated with a dismal prognosis	Colorectal liver metastasis of patients	MMP-12	(101)
Angiopoietin-2	In colorectal lung metastases, the high stromal expression: MMP-1-2,-3 is indicator for a more favorable clinical outcome, whereas high expression of stromal angiopoietin-2 is associated with a reduced cancer-specific survival and an independent prognostic marker for cancer-specific survival in lung metastasis	Colorectal lung metastasis of patients	MMP-1, MMP-2 and MMP-3	(101)
Chemokines related to the immune system and ENPP3, BNIP3, AZGP1 and PIGR	Stage II colorectal cancer. Poor prognosis is associated with low expression of the genes PIGR, CXCL13, MMP3, TUBA1B, CXCL10, and high expression of SESN1, AZGP1, KLK6, EPHA7, SEMA3A, DSC3 ENPP3, BNIP3 and ENPP3	Human Stage II colorectal cancer	MMP3	(102)
MIF	Increased expressions of both MIF and MMP-9 were significantly associated with microvessel density of tumor, but only dual high-expression of MIF and MMP-9 was in relation to tumor invasion and tumor recurrence	67 intracranial meningioma from patients; 57 benign tumors (WHO grade I) and 10 non-benign (WHO grade II/III)	MMP-9	(103)
TGF- β	TGF- β -pretreated A549 cells increased migration and invasiveness, decreased expression of E-cadherin, tight-junction proteins and increased expression of N-cadherin and vimentin. TGF- β -mediated exosomes and might function by increasing the expression of MMP-2	Human Carcinoma lung A549 cell line	MMP-2	(104)
TGF- β , IL-1 α	Production of IL-1 α by pancreatic stellate cells induce alterations in MMP and TIMP profiles and activities, upregulating MMP-1 and MMP-3. TGF- β counteracted the effects of IL-1 α on pancreatic stellate cells downregulating and reestablishing MMP and TIMP profiles	Pancreatic Stellate Cells from patients with pancreatic ductal adenocarcinoma	MMP-1 and MMP-3	(105)
EGR1	EGR1 mediates hypoxia-induced SIRT1 transcriptional repression, and the acetylation of NF- κ B and the activation of MMP-2 and MMP-9	Human HCT 116 and SW480 Cell colorectal cancer cells line	MMP-2 and MMP-9	(106)
CHI3L1 (Chitinase 3-like protein 1)	CHI3L1 promotes the metastasis of gastric and breast cancer cells, interacts with the IL-13R α receptor on the plasma membrane of gastric cancer cells. Even more, CHI3L1 activates MAPK signaling pathway in gastric and breast cancers and the activator protein-1 (AP) transcriptional activity in cancer cells	Gastric cancer cells: MKN-45, AGS, MGC-803 and HGC-27. Breast cancer cells: MDA-MB-231, MDA-MB-435 and MDA-MB-468. Melanoma cells (A375)	MMP-1, MMP-2, MMP-3, MMP-7, MMP-9, MMP-12	(107)



the contribution of MMP-2 and MMP-9 to cancer angiogenesis through the degradation of ECM components and the activation of pro-angiogenic factors VEGF and TGF- β in diverse cancer tissues (134). The above findings may explain the central role of the metalloproteinases MMP-2 and MMP-9 in tumor angiogenesis through the induction of pro-angiogenic factors.

Many other MMPs have also been implicated in the incipient establishment of cancer angiogenesis (Tables 1, 2). For example, MMP-3 and MMP-7 interact *in vivo* with osteopontin at tumor sites and may be related to the angiogenic process during tumor development (135). The interaction of diverse MMPs with another class of proteases also contributes to tumor angiogenesis. For example, the overexpression of the serine protease matriptase in human carcinoma cells regulates MMP-3 activity, promoting proliferation and angiogenesis of tumor tissues by degradation of surrounding ECM (136). MMP-13 has also been implicated in cancer angiogenesis promotion through tube formation and neo-capillary network development mediated by stimulation of ERK-FAK signaling pathway stimulation. It also stimulates VEGF-A secretion, which contributes to the angiogenic process (137).

It has been widely accepted that MMPs likely play antagonistic roles in regulating cancer angiogenesis. MMP-7 and MMP-9 may be involved in the blockage of cancer angiogenesis by cleaving plasminogen and generating angiostatin molecules

(138). Additionally, cross-talk between MMP-7 and MMP-9 leads to the cleavage of insulin-like growth factor-binding protein 2 (IGFBP-2), an angiogenic activator in major aggressive cancers via the transcriptional regulation of the VEGF gene, showing adverse effects in cancer angiogenesis in some tissues (139, 140). It has also been described that MMP-19 is essential to the development of nasopharyngeal carcinoma due to its tumor suppressive and anti-angiogenic functions which can reduce secreted MMP-2 and VEGF (141).

Membrane-Type Metalloproteinases (MT-MMPs) in Cancer Angiogenesis

Membrane type 1 matrix metalloproteinase (MT1-MMP) is considered a key mediator of cancer progression and metastasis. The overexpression of MT1-MMP in malignant breast cells significantly enhances VEGF production via the Akt and mTOR signaling pathways activated by the MT1-MMP-VEGFR-2-Src complex, which promotes tumor growth and angiogenesis (142, 143). Apparently, a similar mechanism could be involved in glioblastoma angiogenesis (144, 145). Therefore, it is worth noting that this tumor phenotype appears to be associated with the dependence of Akt-mediated signaling pathway, which is stimulated by several angiogenic factors.

On the other hand, the proteolytic cleavage of semaphorin 4D into its soluble form by MT1-MMP provides a novel molecular mechanism to control tumor-induced angiogenesis in HNSCC (146). In addition, cross-talk between MT1-MMP, MMP-2, and laminin-5 γ 2 chain fragments contributes to the vasculogenic mimicry of melanoma cells (147). In contrast, the colorectal cancer cells that shed MT1-MMP-mediated endoglin fragments exhibit an anti-angiogenic effect (148). Other studies have demonstrated that MT1-MMP and membrane type 2 matrix metalloproteinase (MT2-MMP) work cooperatively as pro-invasive factors that directly lead to Snail1-triggered cell participation in cancer angiogenesis and metastasis (149). In addition, MT2-MMP is a potential EMT mediator in carcinomas that can degrade adherents and tight junction proteins (150). It has been reported that both MT1-MMP and membrane type 3 matrix metalloproteinase (MT3-MMP) modulate pro-MMP-2 activation, whose angiogenic role in cancer was mentioned above, through inhibition by TIMP-2 and TIMP-3 (151). These data show that a cooperative effect of MT-MMPs during cancer angiogenesis is required together with the angiogenic factors.

Moreover, MT4-MMP expression correlates with EGFR activation, which triggers an angiogenic switch through its catalytic activity and induces the dissemination of cancer cells by disturbing the vessel integrity of the primary breast tumor and promoting hematogenous but not lymphatic metastasis (152–154). Finally, it has been shown that a high MT6-MMP expression in cancer cells is associated with tumor growth; however, further experiments are necessary to determine the exact role of this MT-MMP in the angiogenic process (155).

MMPs and the Immune System in Cancer

It is known that uncontrolled angiogenesis, anomalous ECM turnover, decreased growth, and cell migration, as well as inflammatory response, are the result of an imbalance between MMPs activity and their inhibitors, which may be associated with different diseases. Several specific signals are responsible for coordinating the formation, growth, remodeling, and stabilization of blood vessels. It is recognized that excessive growth-promoting signal cues lead to pathological angiogenesis and cancer (15, 156).

In the tumoral microenvironment, there is a complex and dynamically interacting areas involving stromal cells (fibroblasts, myofibroblasts, neuroendocrine cells and immune cells), blood vessels, lymphatic network, and ECM (157, 158), resulting in a tremendous heterogeneity observed in cancer cells. This condition is because tumor cells express and modulate a broad group of signaling pathways including immune modulatory pathways of cytokines and chemokines, which participate in the progression and establishment of cancer cells (Tables 1, 2) (159–169). In this regard, both cytokines and chemokines induce the expression and activity of MMPs, which in turn allow for the activation of pro-inflammatory signaling pathways as well as the activating receptors, for example, expressed on the surface of T cells and NK cells. Of these molecules act as a powerful mechanism to regulate the immune response. We have analyzed and highlighted several studies showing the involvement of

MMPs and their interactions with immune system proteins in angiogenesis and cancer processes as shown in Tables 1, 2.

The presence of secreted extra-cellular vesicles (exosomes) has recently gained importance within the tumor microenvironment (170–172). Exosomes are specific bearers of multiple modulating molecules, such as the antigens for cluster of differentiation (CD), cytokines and chemokines, growth factors (EGF, FGF, IGF), adhesion proteins (L1/CD171), nucleotides (non-coding RNA, miRNA), and metalloproteinases. Exosomes activate and modify the activity of diverse proteins such as immune proteins and receptor ligands into the circulation by proteolytic cleavage, playing a role as effectors and regulators to promote crosstalk between cancer and stromal cells (53, 157, 173–176). In addition, *in vitro* studies revealed that a type of TGF- β mediated exosome derived from lung cancer cells increase the expression of MMP-2 (104), while immunosuppressive exosome secretion from lymphoblastoid cells induces apoptosis in CD4⁺ T cells (177). Therefore, exosomes could function as conversion markers of malignant cells.

Since they are diverse, MMPs influence multiple cellular processes, such as the inflammatory process regulating barrier function and the activity of inflammatory cytokines and chemokines. Chemoattractant proteins such as MCP-1, MCP-2, MCP-3, and MCP-4 are targets for MMP activity, as result the modified MCPs changing their activity from agonist to antagonist and causing inflammation. Inflammation produces immune tolerance and leads to specific micro-environment conditions, exploited by tumors to evade immune cells and enhance progression, angiogenesis and metastasis (78, 171). In the inflammation process, FGF2 expression can facilitate induction of FGF-dependent angiogenesis by mononuclear phagocytes, T-lymphocytes, and mast cells. The mechanism mediated through FGF2 release induces pro-inflammatory molecules, such as IFN- α , IL-2, IL1- β , and nitric oxide. During the first phases of the EC angiogenesis process, FGFs (1, 2, and 4) upregulate urokinase-type plasminogen activator (uPA) *in vitro* and transform plasmin into plasminogen, an activator of MMPs, triggering ECM degradation and the secretion of exosomes containing MMP-2, MMP-9, TIMP-1, and TIMP-2 (156, 178).

Several studies, both clinical and experimental, have shown that elevated MMP (including MT1-MMPs) levels are associated with the modulation of tumor progression. In brain tumors, growth factors and cytokines modulate the activity of several MMPs. Additionally, it has been observed that MMP-2-positive tumor cells in patients are correlated with low mean survival (54, 179, 180).

Furthermore, in the context of immune cells, there are tumor-associated cells that contribute to the synthesis and upregulation of MMPs (70, 181–183). Importantly, tumor-associated macrophages (TAMs) secrete membrane-bound or soluble proteases, such as MMP-2, MMP-9, and MMP-12, which are involved in ECM degradation and promote the infiltration of tumor-associated blood vessels (184). Macrophages are known to promote cancer initiation and tumor development in an inflammatory environment (185). Bone marrow-derived myeloid cells are also involved in the process, through active regulation of blood vessel formation and maintenance in tumors (186).

Furthermore, accumulated evidence shows that primary tumors can recruit immune cells, such as MMP-9 positive neutrophils, B cells, and M2 polarized macrophages to produce tumor-associated immune cells, which are known to contribute to neovascularization by supplying MMP-9 and other MMPs (59–61, 90, 183, 187). Although most of the published studies consider that M2 macrophages produce high amounts of IL-10, IL-1 β , VEGF and MMP, additional subsets have been described with different proportions of cytokines related to cancer microenvironment. A subset of high M1 and low M2 infiltration macrophages are associated with improved patient survival in non-small-cell lung cancer (188), while the activation of M2 macrophages is correlated with a negative prognosis in cancer progression (189). It is therefore important to reorient the associated functions of the M2 macrophage subset to stop and kill cancer cells.

Finally, the molecular role of MMPs in the immune system and cancer is to modulate a series of latent signaling proteins located in ECM, including cytokines and growth factors such as quiescent TGF- β forming a complex with TGF- β -binding protein-1 in ECM. Thus, TGF- β modulates MMP expression, resulting in a bidirectional regulatory loop enhancing TGF- β signaling and promoting cancer progression (133, 190–194). Another mechanism observed is MMP-9 activity, which truncates IL-8 (1–19, 21, 22, 26–52, 63, 108–135) into more active chains, altering the function of the receptor and improving its biological activity, resulting in greater chemotaxis for neutrophils than the intact form of the cytokine (195, 196).

Accordingly, immunomodulatory mechanisms of MMPs, cytokines, receptors, and growth drivers are involved in the development and progression of several types of cancer.

THERAPEUTIC PERSPECTIVE OF MMPS

MMPs and their inhibitors TIMP, control a wide variety of physiological processes. They constitute promising pharmaceutical targets for inhibition and other metastatic processes.

Currently, monoclonal antibodies are possible candidates to inhibit the activity of MMPs (MMP-14, -12, -9, and -2). However, studies have only managed to identify antibodies against MMP-9 activity, which has biological functions and not for the MMP-14, -12, and -2 (197). In prostate cancer, MMP-9 may amplify local angiogenesis by cleaving membrane-bound VEGF. Therefore, VEGF is a candidate to be blocked and controlled and to prevent the activation of the androgen receptor (AR)/phosphatidylinositol 4-phosphate 5-kinase type-1 alpha (PIP5K1 α)/AKT/MMP-9/VEGF signaling axis required for cell survival and invasion of metastatic tumors (198). Similarly, the effect of phytochemicals on MMP-2, MMP-9, and their tissue inhibitors (TIMPs) has been tested in breast cancer, with no alterations observed *in vitro* (199). TIMP-3 has been another important target of study regarding the inhibition of cancer cell migration, invasion, and metastasis *in vitro* and *in vivo* by natural products (200).

Recently, the effect of MMP-2 gene silencing in normal and MCF-7 cells exposed to the irradiation has been studied. It is known that this MMP leads to the degradation of basement membranes; however, the differential response to DNA damage silencing the MMP-2 gene in normal and MCF-7 cells may be attributable to ROS generation (201). In addition, thrombospondin-2 (THBS2) is a target gene of microRNA-93-5p (miR-93-5p) and THBS2 is closely associated with ECM and MMP-2 and 9. This MicroRNA is involved in the progression of malignant tumors and is highly expressed in cervical cancer tissues and cells. Thus, the THBS2/MMP signaling pathway is relevant for more studies in clinical trials (202). Moreover, the combined therapy for glioma treatment using temozolomide-marimast (a specific alkylating agent and an MMP inhibitor, respectively) results in tumor cell progression and invasiveness. An alternative treatment proposed in an *in vitro* study uses a combination of temozolomide and compounds 1 and 2 of N-O-isopropyl sulfonamido-based hydroxamates (MMP-2 inhibitors) to inhibit cancer cell invasiveness and viability (203).

All these studies represent advances in cancer drug development and cancer therapy, with a focus on the control of MMPs and the proteins with which they form complex networks of multifunctional interactions to modulate the signaling pathways that deviate during the development of metastatic cancer. Importantly, the emerging combined clinical therapy mitigates the side effects of existing treatments and raises the anticancer efficacy of chemotherapeutic drugs.

CONCLUSION REMARKS

In this work, we have highlighted the role that MMPs play in the cancer and its interaction with growth factors, inhibitor proteins, and the EMT process. The activity of MMPs is involved in the degradation, remodeling, and exchange of ECM, which, under normal physiological conditions, contributes to homeostasis as part of an extensive network of extracellular tissue modulation. In cancer, homeostasis is modified, leading to localized abnormal physiological conditions that modify this extensive network of extracellular tissue modulation.

The increase in MMP activities, as an abnormal process, is a way of producing/inducing an erroneous metabolic cascade. Erroneous metabolic cascades are signals that trigger the emergence of complex abnormal cell pathways, which give rise to tumor/cancerous phenotype cells. In this regard, the transformation into tumor/cancerous phenotypes suggests an exacerbated adaptive survival process. MMPs are not the only elements of this extensive network of extracellular tissue modulation; others such as TIMP proteins, which modulate MMPs, ADAMs and ADAMTSs (15), play a role in this anomalous process as numerous regulatory branches of the network do, namely interleukins.

Although the information on the role of MMPs in cancer is very broad and the way these proteins are expressed is well known, we observed a significant lack of data at the fine level of the relationship between MMPs and the continuity of both the normal and altered signals that positively modulate the

carcinogenic process. MMPs produce modulatory elements that remain unclear and, considering that the ECM is a complex array of proteins, fibers, and carbohydrates in different tissues, there may be several variants that generate the loss of homeostasis, causing the diverse cancerous processes observed.

In the angiogenesis process, MMPs are well-known key factors involved in ECM degradation that induce angiogenesis initiation in both physiological and pathological processes. However, the experimental evidence thus far demonstrates that MMPs also play a decisive role in the activation of pro-angiogenic and, in some cases, anti-angiogenic factors in cancer tissues. Thus, MMPs can be considered angiomodulators, which could control new vessel formation necessary for cancer growth, progression, and spread. Therefore, we speculate that MMPs participate in cancer angiogenesis in a cell context-dependent manner.

Most of the experimental data regarding MMP participation in cancer development, vascular endothelium processes, other epithelia (such as periodontal), and inflammatory processes allow us to assume that MMPs are proteins that carry out a type of external cellular regulation/signaling on the ECM. These proteins possess a different regulatory action mechanism that is complementary to other mechanisms such as ligand-receptor signaling pathways. The MMP mechanism is based on editing macromolecules by proteolysis, mainly anchored to the ECM.

It is evident that the proportion of MMPs and other macromolecules (cytokines, growth factors, fibers such as integrins, polysaccharides, and others) in the ECMs of different tissues in normal conditions are metabolically, micro-environmentally, and epigenetically balanced for their functions in each type of tissue. There are tissue-specific proportions, although ECMs have a high degree of heterogeneity.

Inspecting our concept of MMP participation in ECM in the literature, we found an excellent review of ECM with similar concepts (204). The ECM is a highly dynamic system in constant remodeling and is undoubtedly an extension of communication/modulation/signaling among cells located outside the plasma membrane, where MMPs participate in protein editing by providing post-translational modifications.

On the other hand, EMT is a biological process aimed producing mesenchymal phenotype cells from epithelial cells. Its inverse process, mesenchymal-epithelial transition (MET), is carried out with the participation of ECM elements. EMT and MET lead to normal tissue regeneration and fibrosis [EMT type 2, according to Kalluri (205)]. Regarding the participation of MMPs in EMT type 3 (abnormal type), it is evident that their participation is associated with errors in the communication,

modulation, and signaling of this process, which is induced and directed by tumor cells.

Evidence suggests that tumor cells induce the uncontrolled upregulation of MMPs, producing a large number of stimulating factors that disrupt EMT and immunological processes that prevent tumor cell elimination and migration. The upregulation progresses to generating anomalous tissue-specific type signals.

It is known that tumor cells have extensive heterogeneity in their metabolism and phenotype relative to normal tissue across cancer types. Furthermore, these abnormal signals coming from the tumor cells are tissue-specific, leading to the adaptation to the microenvironment where they developed. Finally, an interesting observation of the MMP family is the large, robust specificity profile, which suggests that its role is controlled in a tissue-specific manner; that is, MMP types are expressed accordingly to the regulatory proteins needed for the tissue.

However, more research efforts are needed to determine when abnormal signals begin, what the determinants are, and how microenvironmental tissue-specific conditions can lead a cell to change its metabolism and phenotype. In addition, the question remains: When does the high expression problem of MMPs become a problem metabolically? Although MMPs do not seem to be the cause of the appearance of tumor cells, they induce tumor development because they are targets to regulate development, contributing to increased invasiveness and growth of metastatic tumors.

AUTHOR CONTRIBUTIONS

MA-S conceived the idea and scripted the basis of the manuscript. RA and MA-S had equal contributions and a role in the design, analysis, and writing of the article. RA contributed in entirety to the design of the **Figure 1**. SQ-F conceived the idea. SQ-F and RA designed of the **Figure 2**, **Table 2**, and drafted the manuscript. RA conceived and designed the **Figure 3B** and participated with JT-R, VA-A, JL-R, MR-C, and MA-S in the designed the **Figure 3A**. EB-V participated in its construction. All authors reviewed the literature, critically reviewed the manuscript, and approved the final version.

FUNDING

This present study was supported by the Autonomous University of Mexico City (UACM) and by National Institute of Psychiatry Ramón de la Fuente Muñiz (INPRFM).

REFERENCES

- Di Cara G, Marabeti M, Musso R, Riili I, Cancemi P, Pucci Minafra I. New insights into the occurrence of matrix metalloproteinases-2 and-9 in a Cohort of Breast Cancer Patients and Proteomic Correlations. *Cells*. (2018) 7:89. doi: 10.3390/cells7080089
- Valastyan S, Weinberg RA. Tumor metastasis: molecular insights and evolving paradigms. *Cell*. (2011) 147:275–92. doi: 10.1016/j.cell.2011.09.024
- Seyfried TN, Huysentruyt LC. On the origin of cancer metastasis. *Crit Rev Oncog*. (2013) 18:43–73. doi: 10.1615/CritRevOncog.v18.i1-2.40
- Thiery JP. Epithelial–mesenchymal transitions in tumour progression. *Nat Rev Cancer*. (2002) 2:442–54. doi: 10.1038/nrc822
- Yu D, Ye T, Xiang Y, Shi Z, Zhang J, Lou B, et al. Quercetin inhibits epithelial–mesenchymal transition, decreases invasiveness and metastasis, and reverses IL-6 induced epithelial–mesenchymal transition, expression of MMP by inhibiting STAT3 signaling in pancreatic cancer cells. *Oncol Targets Ther*. (2017) 10:4719–29. doi: 10.2147/OTT.S136840

6. Horejs CM. Basement membrane fragments in the context of the epithelial-to-mesenchymal transition. *Eur J Cell Biol.* (2016) 95:427–40. doi: 10.1016/j.ejcb.2016.06.002
7. Ehrmann RL, Knoth M. Choriocarcinoma: transfilter stimulation of vasoproliferation in the hamster cheek pouch—studied by light and electron microscopy. *J Natl Cancer Inst.* (1968) 41:1329–41.
8. Bielenberg DR, Zetter BR. The Contribution of Angiogenesis to the Process of Metastasis. *Cancer J.* (2015) 21:267–73. doi: 10.1097/PPO.0000000000001138
9. Fukushima R, Kasamatsu A, Nakashima D, Higo M, Fushimi K, Kasama H, et al. Overexpression of translocation associated membrane protein 2 leading to cancer-associated matrix metalloproteinase activation as a putative metastatic factor for human oral cancer. *J Cancer.* (2018) 9:3326–33. doi: 10.7150/jca.25666
10. Miyake M, Goodison S, Lawton A, Gomes-Giacoa E, Rosser CJ. Angiogenin promotes tumoral growth and angiogenesis by regulating matrix metalloproteinase-2 expression via the ERK1/2 pathway. *Oncogene.* (2015) 34:890–901. doi: 10.1038/onc.2014.2
11. Saito T, Kasamatsu A, Ogawara K, Miyamoto I, Saito K, Iyoda M, et al. Semaphorin7A promotion of tumoral growth and metastasis in human oral cancer by regulation of G1 cell cycle and matrix metalloproteinases: possible contribution to tumoral angiogenesis. *PLoS ONE.* (2015) 10:e0137923. doi: 10.1371/journal.pone.0137923
12. Yamamoto A, Kasamatsu A, Ishige S, Koike K, Saito K, Kouzu Y, et al. Exocyst complex component Sec8: a presumed component in the progression of human oral squamous-cell carcinoma by secretion of matrix metalloproteinases. *J Cancer Res Clin Oncol.* (2013) 139:533–42. doi: 10.1007/s00432-012-1356-2
13. Mook ORF, Frederiks WM, Van Noorden CJF. The role of gelatinases in colorectal cancer progression and metastasis. *Biochim Biophys Acta Rev Cancer.* (2004) 1705:69–89. doi: 10.1016/j.bbcan.2004.09.006
14. Nagase H, Visse R, Murphy G. Structure and function of matrix metalloproteinases and TIMPs. *Cardiovasc Res.* (2006) 69:562–73. doi: 10.1016/j.cardiores.2005.12.002
15. Brew K, Nagase H. The tissue inhibitors of metalloproteinases (TIMPs): an ancient family with structural and functional diversity. *Biochim Biophys Acta.* (2010) 1803:55–71. doi: 10.1016/j.bbamcr.2010.01.003
16. Kelwick R, Desanlis I, Wheeler GN, Edwards DR. The ADAMTS (A Disintegrin and Metalloproteinase with Thrombospondin motifs) family. *Genome Biol.* (2015) 16:113. doi: 10.1186/s13059-015-0676-3
17. Gross, Jerome and Lapiere C. Collagenolytic activity in amphibian tissues: a tissue culture assay. *Pnas.* (1962) 48:1014–22. doi: 10.1073/pnas.48.6.1014
18. Chen Q, Jin M, Yang F, Zhu J, Xiao Q, Zhang L. Matrix metalloproteinases: inflammatory regulators of cell behaviors in vascular formation and remodeling. *Mediators Inflamm.* (2013) 2013:928315. doi: 10.1155/2013/928315
19. Wang X, Khalil RA. Chapter 8: Matrix metalloproteinases, vascular remodeling, and vascular disease. In: Khalil RA, editor. *Advances in Pharmacology.* San Diego, CA: Academic Press (2018). p. 241–330. doi: 10.1016/bs.apha.2017.08.002
20. Humphrey W, Dalke A, Schulten K. VMD: Visual molecular dynamics. *J Mol Graph.* (1996) 14: 33–8. doi: 10.1016/0263-7855(96)00018-5
21. Murphy G, Nagase H. Progress in matrix metalloproteinase research. *Mol Aspects Med.* (2008) 29:290–308. doi: 10.1016/j.mam.2008.05.002
22. Jabłonska-Trypuć A, Matejczyk M, Rosochacki S. Matrix metalloproteinases (MMPs), the main extracellular matrix (ECM) enzymes in collagen degradation, as a target for anticancer drugs. *J Enzyme Inhib Med Chem.* (2016) 31:177–83. doi: 10.3109/14756366.2016.1161620
23. Kujawski M, Kay H, Yu H, Kujawski M, Kortylewski M, Lee H, et al. angiogenesis in mice Find the latest version : Stat3 mediates myeloid cell – dependent tumor angiogenesis in mice. (2008) 118:3367–77. doi: 10.1172/JCI35213
24. Lee EJ, Park SS, Kim WJ, Moon SK. IL-5-induced migration via ERK1/2-mediated MMP-9 expression by inducing activation of NF- κ B in HT1376 cells. *Oncol Rep.* (2012) 28:1084–90. doi: 10.3892/or.2012.1857
25. Lee EJ, Lee SJ, Kim S, Cho SC, Choi YH, Kim WJ, et al. Interleukin-5 enhances the migration and invasion of bladder cancer cells via ERK1/2-mediated MMP-9/NF- κ B/AP-1 pathway: Involvement of the p21WAF1 expression. *Cell Signal.* (2013) 25:2025–38. doi: 10.1016/j.cellsig.2013.06.004
26. Eckhard U, Huesgen PF, Schilling O, Bellac CL, Butler GS, Cox JH, et al. Active site specificity profiling of the matrix metalloproteinase family: proteomic identification of 4300 cleavage sites by nine MMPs explored with structural and synthetic peptide cleavage analyses. *Matrix Biol.* (2016) 49:37–60. doi: 10.1016/j.matbio.2015.09.003
27. Kridel SJ, Chen E, Kotra LP, Howard EW, Mobashery S, Smith JW. Substrate hydrolysis by matrix metalloproteinase-9. *J Biol Chem.* (2001) 276:20572–8. doi: 10.1074/jbc.M100900200
28. Gomiz-Rüth FX. Catalytic domain architecture of metzincin metalloproteinases. *J Biol Chem.* (2009) 284:15353–7. doi: 10.1074/jbc.R800069200
29. Tallant C, Marrero A, Gomiz-Rüth FX. Matrix metalloproteinases: Fold and function of their catalytic domains. *Biochim Biophys Acta Mol Cell Res.* (2010) 1803:20–8. doi: 10.1016/j.bbamcr.2009.04.003
30. Visse R, Nagase H. Matrix metalloproteinases and tissue inhibitors of metalloproteinases. *Circ Res.* (2003) 92:827–39. doi: 10.1161/01.RES.0000070112.80711.3D
31. Rajabi M, Mousa S. The role of angiogenesis in cancer treatment. *Biomedicine.* (2017) 5:34. doi: 10.3390/biomedicine5020034
32. Rundhaug JE. Matrix metalloproteinases and angiogenesis Angiogenesis Review Series. *J Cell Mol Med.* (2005) 9:267–85. doi: 10.1111/j.1582-4934.2005.tb00355.x
33. Carmeliet P, Jain RK. Molecular mechanisms and clinical applications of angiogenesis. *Nature.* (2011) 473:298–307. doi: 10.1038/nature10144
34. Sang QX. MINIREVIEW Complex role of matrix metalloproteinases in angiogen- esis. *Cell Res.* (1998) 8:171–7. doi: 10.1038/cr.1998.17
35. Stetler-Stevenson WG. Matrix metalloproteinases in angiogenesis: a moving target for therapeutic intervention. *J Clin Invest.* (1999) 103:1237–41. doi: 10.1172/JCI6870
36. Smith SK. Angiogenesis. *Semin Reprod Med.* (1997) 15:221–7. doi: 10.1055/s-2008-1068751
37. Benedito R, Hellström M. Notch as a hub for signaling in angiogenesis. *Exp Cell Res.* (2013) 319:1281–8. doi: 10.1016/j.yexcr.2013.01.010
38. Chambers AF, Groom AC, MacDonald IC. Dissemination and growth of cancer cells in metastatic sites. *Nat Rev Cancer.* (2002) 2:563–72. doi: 10.1038/nrc865
39. Paduch R. The role of lymphangiogenesis and angiogenesis in tumor metastasis. *Cell Oncol.* (2016) 39:397–410. doi: 10.1007/s13402-016-0281-9
40. Alitalo A, Detmar M. Interaction of tumor cells and lymphatic vessels in cancer progression. *Oncogene.* (2012) 31:4499–508. doi: 10.1038/ncr.2011.602
41. Sleeman JP, Thiele W. Tumor metastasis and the lymphatic vasculature. *Int J Cancer.* (2009) 125:2747–56. doi: 10.1002/ijc.24702
42. Burton JB, Priceman SJ, Sung JL, Brakenhielm E, Dong SA, Pytowski B, et al. Suppression of prostate cancer nodal and systemic metastasis by blockade of the lymphangiogenic axis. *Cancer Res.* (2008) 68:7828–37. doi: 10.1158/0008-5472.CAN-08-1488
43. Davydova N, Harris NC, Roufail S, Paquet-Fifield S, Ishaq M, Streltsov VA, et al. Differential receptor binding and regulatory mechanisms for the lymphangiogenic growth factors vascular endothelial growth factor (VEGF)-C and -D. *J Biol Chem.* (2016) 291:27265–78. doi: 10.1074/jbc.M116.736801
44. Roberts N, Kloos B, Cassella M, Podgrabska S, Persaud K, Wu Y, et al. Inhibition of VEGFR-3 activation with the antagonistic antibody more potently suppresses lymph node and distant metastases than inactivation of VEGFR-2. *Cancer Res.* (2006) 66:2650–57. doi: 10.1158/0008-5472.CAN-05-1843
45. Aldair T, Montani J. Chapter 1: Overview of angiogenesis. In: Granger N, Granger J, editors. *Angiogenesis.* San Rafael, CA: Morgan & Claypool Life Sciences (2010). p. 1–84. doi: 10.4199/C00017ED1V01Y201009 ISP010
46. Jadvar H. Chapter 12: Angiogenesis. In: Jadvar H, Jacene H, Graham M, editors. *Molecular Imaging: An Introduction.* Los Angeles, CA: Cambridge University Press (2017). p. 52–4. doi: 10.1017/9781107360044.013
47. Prager GW, Zielinski CC. Angiogenesis in cancer. In: Jawahar LM, Dhalla NS, editors. *Biochemical Basis and Therapeutic Implications*

- of *Angiogenesis*. New York, NY: Springer (2017). p. 335–56. doi: 10.1007/978-1-4614-5857-9_18
48. Semenza GL. Targeting HIF-1 for cancer therapy. *Nat Rev Cancer*. (2003) 3:721–32. doi: 10.1038/nrc1187
 49. Adams RH, Alitalo K. Molecular regulation of angiogenesis and lymphangiogenesis. *Nat Rev Mol Cell Biol*. (2007) 8:464–78. doi: 10.1038/nrm2183
 50. Shibuya M, Claesson-Welsh L. Signal transduction by VEGF receptors in regulation of angiogenesis and lymphangiogenesis. *Exp Cell Res*. (2006) 312:549–60. doi: 10.1016/j.yexcr.2005.11.012
 51. Egeblad M, Nakasone ES, Werb Z. Tumors as organs: complex tissues that interface with the entire organism. *Dev Cell*. (2010) 18:884–901. doi: 10.1016/j.devcel.2010.05.012
 52. Majumder P. Integrin-mediated delivery of drugs and nucleic acids for anti-angiogenic cancer therapy: current landscape and remaining challenges. *Bioengineering*. (2018) 5:76. doi: 10.3390/bioengineering5040076
 53. Park JE, Tan H Sen, Datta A, Lai RC, Zhang H, Meng W, et al. Hypoxic tumor cell modulates its microenvironment to enhance angiogenic and metastatic potential by secretion of proteins and exosomes. *Mol Cell Proteomics*. (2010) 9:1085–99. doi: 10.1074/mcp.M900381-MCP200
 54. Roomi MW, Kalinovsky T, Rath M, Niedzwiecki A. Modulation of MMP-2 and MMP-9 secretion by cytokines, inducers and inhibitors in human glioblastoma T-98G cells. *Oncol Rep*. (2017) 37:1907–13. doi: 10.3892/or.2017.5391
 55. Huang Q, Lan F, Wang X, Yu Y, Ouyang X, Zheng F, et al. IL-1 β -induced activation of p38 promotes metastasis in gastric adenocarcinoma via upregulation of AP-1/c-fos, MMP2 and MMP9. *Mol Cancer*. (2014) 13:1–15. doi: 10.1186/1476-4598-13-18
 56. Ji Y, Yang X, Li J, Lu Z, Li X, Yu J, et al. IL-22 promotes the migration and invasion of gastric cancer cells via IL-22R1/AKT/MMP-9 signaling. *Int J Clin Exp Pathol*. (2014) 7:3694–703. Available online at: <http://www.ijcep.com/files/ijcep0000514.pdf>
 57. Fukui H, Zhang X, Sun C, Hara K, Kikuchi S, Yamasaki T, et al. IL-22 produced by cancer-associated fibroblasts promotes gastric cancer cell invasion via STAT3 and ERK signaling. *Br J Cancer*. (2014) 111:763–71. doi: 10.1038/bjc.2014.336
 58. Cardoso AP, Pinto ML, Pinto AT, Monteiro C, Oliveira MI, et al. Matrix metalloproteinases as maestros for the dual role of LPS- and IL-10-stimulated macrophages in cancer cell behaviour. *BMC Cancer*. (2015) 15:1–14. doi: 10.1186/s12885-015-1466-8
 59. Espinoza-Sánchez NA, Chimal-Ramírez GK, Mantilla A, Fuentes-Pananá EM. IL-1 β , IL-8, and matrix metalloproteinases-1, -2, and -10 are enriched upon monocyte-breast cancer cell cocultivation in a matrigel-based three-dimensional system. *Front Immunol*. (2017) 8:205. doi: 10.3389/fimmu.2017.00205
 60. Ning Y, Cui Y, Li X, Cao X, Chen A, Xu C, et al. Co-culture of ovarian cancer stem-like cells with macrophages induced SKOV3 cells stemness via IL-8/STAT3 signaling. *Biomed Pharmacother*. (2018) 103:262–71. doi: 10.1016/j.biopha.2018.04.022
 61. Ou Z, Wang Y, Liu L, Li L, Yeh S, Qi L, et al. Tumor microenvironment B cells increase bladder cancer metastasis via modulation of the IL-8/androgen receptor (AR)/MMPs signals. *Oncotarget*. (2015) 6:26065–78. doi: 10.18632/oncotarget.4569
 62. Ha NH, Park DG, Woo BH, Kim DJ, Choi J Il, Park BS, et al. Porphyromonas gingivalis increases the invasiveness of oral cancer cells by upregulating IL-8 and MMPs. *Cytokine*. (2016) 86:64–72. doi: 10.1016/j.cyto.2016.07.013
 63. Li A, Dubey S, Varney ML, Dave BJ, Singh RK. IL-8 Directly enhanced endothelial cell survival, proliferation, and matrix metalloproteinases production and regulated angiogenesis. *J Immunol*. (2003) 170:3369–76. doi: 10.4049/jimmunol.170.6.3369
 64. Epanchintsev A, Shyamsunder P, Verma RS, Lyakhovich A. IL-6, IL-8, MMP-2, MMP-9 are overexpressed in Fanconi anemia cells through a NF- κ B/TNF- α dependent mechanism. *Mol Carcinog*. (2015) 54:1686–99. doi: 10.1002/mc.22240
 65. Desai S, Laskar S, Pandey BN. Autocrine IL-8 and VEGF mediate epithelial-mesenchymal transition and invasiveness via p38/JNK-ATF-2 signalling in A549 lung cancer cells. *Cell Signal*. (2013) 25:1780–91. doi: 10.1016/j.cellsig.2013.05.025
 66. Bruno A, Bassani B, D'Urso DG, Pitaku I, Cassinotti E, Pelosi G, et al. Angiogenin and the MMP9-TIMP2 axis are up-regulated in proangiogenic, decidual NK-like cells from patients with colorectal cancer. *FASEB J*. (2018) 32:5365–77. doi: 10.1096/fj.201701103R
 67. Bates AM, Gomez Hernandez MP, Lanzel EA, Qian F, Brogden KA. Matrix metalloproteinase (MMP) and immunosuppressive biomarker profiles of seven head and neck squamous cell carcinoma (HNSCC) cell lines. *Transl Cancer Res*. (2018) 7:533–42. doi: 10.21037/tcr.2018.05.09
 68. Kim SJ, Pham TH, Bak Y, Ryu HW, Oh SR, Yoon DY. Orientin inhibits invasion by suppressing MMP-9 and IL-8 expression via the PKC α /ERK/AP-1/STAT3-mediated signaling pathways in TPA-treated MCF-7 breast cancer cells. *Phytomedicine*. (2018) 50:35–42. doi: 10.1016/j.phymed.2018.09.172
 69. Jayatilaka H, Umanson FG, Shah V, Meirson T, Russo G, Starich B, et al. Tumor cell density regulates matrix metalloproteinases for enhanced migration. *Oncotarget*. (2018) 9:32556–69. doi: 10.18632/oncotarget.25863
 70. Zhang Y, Wang S, Liu Z, Yang L, Liu J, Xiu M. Increased Six1 expression in macrophages promotes hepatocellular carcinoma growth and invasion by regulating MMP-9. *J Cell Mol Med*. (2019) 23:4523–33. doi: 10.1111/jcmm.14342
 71. Cutler SJ, Doecke JD, Ghazawi I, Yang J, Griffiths LR, Spring KJ, et al. Novel STAT binding elements mediate IL-6 regulation of MMP-1 and MMP-3. *Sci Rep*. (2017) 7:1–12. doi: 10.1038/s41598-017-08581-y
 72. Zergoun AA, Zebboudj A, Sellam SL, Kariche N, Djennaoui D, Ouraghi S, et al. IL-6/NOS2 inflammatory signals regulate MMP-9 and MMP-2 activity and disease outcome in nasopharyngeal carcinoma patients. *Tumor Biol*. (2016) 37:3505–14. doi: 10.1007/s13277-015-4186-4
 73. Chen W, Xia T, Wang D, Huang B, Zhao P, Wang J, et al. Human astrocytes secrete IL-6 to promote glioma migration and invasion through upregulation of cytomembrane MMP14. *Oncotarget*. (2016) 7:62425–38. doi: 10.18632/oncotarget.11515
 74. Ernst W, Najdovska M, Grail D, Lundgren-May T, Buchert M, Tye H, et al. STAT3 and STAT1 mediate IL-11-dependent and inflammation-associated gastric tumorigenesis in gp130 receptor mutant mice. *J Clin Invest*. (2008) 118:1727–38. doi: 10.1172/JCI34944
 75. Lim JH. Inhibition of the interleukin-11-STAT3 axis attenuates hypoxia-induced migration and invasion in MDA-MB-231 breast cancer cells. *Korean J Physiol Pharmacol*. (2014) 18:391–6. doi: 10.4196/kjpp.2014.18.5.391
 76. Yue T, Zheng X, Dou Y, Zheng X, Sun R, Tian Z, et al. Interleukin 12 shows a better curative effect on lung cancer than paclitaxel and cisplatin doublet chemotherapy. *BMC Cancer*. (2016) 16:1–13. doi: 10.1186/s12885-016-2701-7
 77. Scott KA, Holdsworth H, Balkwill FR, Dias S. Exploiting changes in the tumour microenvironment with sequential cytokine and matrix metalloprotease inhibitor treatment in a murine breast cancer model. *Br J Cancer*. (2000) 83:1538–43. doi: 10.1054/bjoc.2000.1487
 78. Fabre JAS, Giustinniani J, Garbar C, Merrouche Y, Antonicelli F, Bensussan A. The interleukin-17 family of cytokines in breast cancer. *Int J Mol Sci*. (2018) 19:3880. doi: 10.3390/ijms19123880
 79. Zhu XW, Mulcahy LA, Mohammed RAA, Lee AHS, Franks HA, Kilpatrick L, et al. IL-17 expression by breast-cancer-associated macrophages: IL-17 promotes invasiveness of breast cancer cell lines. *Breast Cancer Res*. (2008) 10:1–11. doi: 10.1186/bcr2195
 80. Amara S, Majors C, Roy B, Hill S, Rose KL, Myles EL, et al. Critical role of SIK3 in mediating high salt and IL-17 synergy leading to breast cancer cell proliferation. *PLoS ONE*. (2017) 12:e180097. doi: 10.1371/journal.pone.0180097
 81. Zhang Q, Liu S, Parajuli KR, Zhang W, Zhang K, Mo Z, et al. Interleukin-17 promotes prostate cancer via MMP7-induced epithelial-to-mesenchymal transition. *Oncogene*. (2017) 36:687–99. doi: 10.1038/onc.2016.240
 82. Ren L, Xu Y, Liu C, Wang S, Qin G. IL-17RB enhances thyroid cancer cell invasion and metastasis via ERK1/2 pathway-mediated MMP-9 expression. *Mol Immunol*. (2017) 90:126–35. doi: 10.1016/j.molimm.2017.06.034
 83. Liu D, Zhang R, Wu J, Pu Y, Yin X, Cheng Y, et al. Interleukin-17A promotes esophageal adenocarcinoma cell invasiveness through ROS-dependent, NF- κ B-mediated MMP-2/9 activation. *Oncol Rep*. (2017) 37:1779–85. doi: 10.3892/or.2017.5426

84. Kobori T, Hamasaki S, Kitaura A, Yamazaki Y, Nishinaka T, Niwa A, et al. Interleukin-18 amplifies macrophage polarization and morphological alteration, leading to excessive angiogenesis. *Front Immunol.* (2018) 9:334. doi: 10.3389/fimmu.2018.00334
85. Zhou Y, Hu Z, Li N, Jiang R. Interleukin-32 stimulates osteosarcoma cell invasion and motility via AKT pathway-mediated MMP-13 expression. *Int J Mol Med.* (2015) 35:1729–33. doi: 10.3892/ijmm.2015.2159
86. Yang F, Wen M, Pan D, Lin X, Mo J, Dong X, et al. IL-33/ST2 axis regulates vasculogenic mimicry via ERK1/2-MMP-2/9 pathway in melanoma. *Dermatology.* (2019) 235:225–33. doi: 10.1159/000498857
87. Yang Z, Gao X, Wang J, Xu L, Zheng Y, Xu Y. Interleukin-33 enhanced the migration and invasiveness of human lung cancer cells. *Onco Targets Ther.* (2018) 11:843–9. doi: 10.2147/OTT.S155905
88. Maywald RL, Doerner SK, Pastorelli L, De Salvo C, Benton SM, Dawson EP, et al. IL-33 activates tumor stroma to promote intestinal polyposis. *Proc Natl Acad Sci USA.* (2015) 112:E2487–96. doi: 10.1073/pnas.1422445112
89. Zucker S, Vacirca J. Role of matrix metalloproteinases (MMPs) in colorectal cancer. *Cancer Metastasis Rev.* (2004) 23:101–17. doi: 10.1023/A:1025867130437
90. Zou J-M, Qin J, Li Y-C, Wang Y, Li D, Shu Y, et al. IL-35 induces N2 phenotype of neutrophils to promote tumor growth. *Oncotarget.* (2017) 8:33501–14. doi: 10.18632/oncotarget.16819
91. Long J, Guo H, Cui S, Zhang H, Liu X, Li D, et al. IL-35 expression in hepatocellular carcinoma cells is associated with tumor progression. *Oncotarget.* (2016) 7:45678–86. doi: 10.18632/oncotarget.10141
92. Chen YH, Zhou BY, Wu GC, Liao DQ, Li J, Liang SS, et al. Effects of exogenous IL-37 on the biological characteristics of human lung adenocarcinoma A549 cells and the chemotaxis of regulatory T cells. *Cancer Biomarkers.* (2018) 21:661–73. doi: 10.3233/CBM-170732
93. Li M, Lu Y, Xu Y, Wang J, Zhang C, Du Y, et al. Horizontal transfer of exosomal CXCR4 promotes murine hepatocarcinoma cell migration, invasion and lymphangiogenesis. *Gene.* (2018) 676:101–9. doi: 10.1016/j.gene.2018.07.018
94. Lee S, Park S, Cho Y, Park K, Kim E, Jung K, et al. Activation of matrix metalloproteinase-9 by TNF- α in human urinary bladder cancer HT1376 cells.pdf. *Oncol Rep.* (2008) 19:1007–13. doi: 10.3892/or.19.4.1007
95. Tanaka T, Imamura T, Yoneda M, Irie A, Ogi H, Nagata M, et al. Enhancement of active MMP release and invasive activity of lymph node metastatic tongue cancer cells by elevated signaling via the TNF- α -TNFR1-NF- κ B pathway and a possible involvement of angiopoietin-like 4 in lung metastasis. *Int J Oncol.* (2016) 49:1377–84. doi: 10.3892/ijo.2016.3653
96. Xia P, Zhang R, Ge G. C/EBP β mediates TNF- α -induced cancer cell migration by inducing MMP expression dependent on p38 MAPK. *J Cell Biochem.* (2015) 116:2766–77. doi: 10.1002/jcb.25219
97. Ohta K, Naruse T, Ishida Y, Shigeishi H, Nakagawa T, Fukui A, et al. TNF- α -induced IL-6 and MMP-9 expression in immortalized ameloblastoma cell line established by hTERT. *Oral Dis.* (2017) 23:199–209. doi: 10.1111/odi.12594
98. Gutwein P, Stoeck A, Riedle S, Gast D, Runz S, Condon TP, et al. Cleavage of L1 in exosomes and apoptotic membrane vesicles released from ovarian carcinoma cells. *Clin Cancer Res.* (2005) 11:2492–501. doi: 10.1158/1078-0432.CCR-04-1688
99. Mahmoodi F, Akrami H. PlGF knockdown decreases tumorigenicity and stemness properties of spheroid body cells derived from gastric cancer cells. *J Cell Biochem.* (2017) 118:851–9. doi: 10.1002/jcb.25762
100. Winkler F, Kozin S V., Tong RT, Chae S-S, Booth MF, Garkavtsev I, et al. Kinetics of vascular normalization by VEGFR2 blockade governs brain tumor response to radiation. *Cancer Cell.* (2004) 6:553–63. doi: 10.1016/j.ccr.2004.10.011
101. Kahlert C, Pecqueur M, Halama N, Dienemann H, Muley T, Pfanschmidt J, et al. Tumour-site-dependent expression profile of angiogenic factors in tumour-associated stroma of primary colorectal cancer and metastases. *Br J Cancer.* (2014) 110:441–9. doi: 10.1038/bjc.2013.745
102. Ågesen TH, Sveen A, Merok MA, Lind GE, Nesbakken A, Skotheim RI, et al. ColoGuideEx: a robust gene classifier specific for stage II colorectal cancer prognosis. *Gut.* (2012) 61:1560–7. doi: 10.1136/gutjnl-2011-301179
103. Huang Q, Zhao SL, Tian XY, Li B, Li Z. Increased co-expression of macrophage migration inhibitory factor and matrix metalloproteinase 9 is as-associated with tumor recurrence of meningioma. *Int J Med Sci.* (2013) 10:276–85. doi: 10.7150/ijms.5185
104. Wu D-M, Deng S-H, Liu T, Han R, Zhang T, Xu Y. TGF- β -mediated exosomal lnc-MMP2-2 regulates migration and invasion of lung cancer cells to the vasculature by promoting MMP2 expression. *Cancer Med.* (2018) 7:5118–29. doi: 10.1002/cam4.1758
105. Tjomsland V, Pomianowska E, Aasrum M, Sandnes D, Verbeke CS, Gladhaug IP. Profile of MMP and TIMP expression in human pancreatic stellate cells: regulation by IL-1 α and TGF β and implications for migration of pancreatic cancer cells. *Neoplasia.* (2016) 18:447–56. doi: 10.1016/j.neo.2016.06.003
106. Yu S, Zhou R, Yang T, Liu S, Cui Z, Qiao Q, et al. Hypoxia promotes colorectal cancer cell migration and invasion in a SIRT1-dependent manner. *Cancer Cell Int.* (2019) 19:1–12. doi: 10.1186/s12935-019-0819-9
107. Chen Y, Zhang S, Wang Q, Zhang X. Tumor-recruited M2 macrophages promote gastric and breast cancer metastasis via M2 macrophage-secreted CHI3L1 protein. *J Hematol Oncol.* (2017) 10:36. doi: 10.1186/s13045-017-0408-0
108. Mott JD, Werb Z. Regulation of matrix biology by matrix metalloproteinases. *Curr Opin Cell Biol.* (2004) 16:558–64. doi: 10.1016/j.ccb.2004.07.010
109. Schenk S, Quaranta V. Tales from the crypt[ic] sites of the extracellular matrix. *Trends Cell Biol.* (2003) 13:366–75. doi: 10.1016/S0962-8924(03)00129-6
110. Mazor R, Alsaigh T, Shaked H, Altshuler AE, Pocock ES, Kistler EB, et al. Matrix metalloproteinase-1-mediated up-regulation of vascular endothelial growth factor-2 in endothelial cells. *J Biol Chem.* (2013) 288:598–607. doi: 10.1074/jbc.M112.417451
111. Ito TK, Ishii G, Saito S, Yano K, Hoshino A, Suzuki T, et al. Degradation of soluble VEGF receptor-1 by MMP-7 allows VEGF access to endothelial cells. *Blood.* (2009) 113:2363–9. doi: 10.1182/blood-2008-08-172742
112. Hanemaaijer R, Sorsa T, Kontinen YT, Ding Y, Sutinen M, Visser H, et al. Matrix metalloproteinase-8 is expressed in rheumatoid synovial fibroblasts and endothelial cells. *J Biol Chem.* (2002) 272:31504–9. doi: 10.1074/jbc.272.50.31504
113. Cheng XW, Kuzuya M, Nakamura K, Maeda K, Tsuzuki M, Kim W, et al. Mechanisms underlying the impairment of ischemia-induced neovascularization in matrix metalloproteinase 2-deficient mice. *Circ Res.* (2007) 100:904–13. doi: 10.1161/01.RES.0000260801.12916.b5
114. Fang C, Wen G, Zhang L, Lin L, Moore A, Wu S, et al. An important role of matrix metalloproteinase-8 in angiogenesis *in vitro* and *in vivo*. *Cardiovasc Res.* (2013) 99:146–55. doi: 10.1093/cvr/cvt060
115. Tarabozetti G, D'Ascenzo S, Borsotti P, Giavazzi R, Pavan A, Dolo V. Shedding of the matrix metalloproteinases MMP-2, MMP-9, and MT1-MMP as membrane vesicle-associated components by endothelial cells. *Am J Pathol.* (2002) 160:673–80. doi: 10.1016/S0002-9440(10)64887-0
116. Hamano Y, Zeisberg M, Sugimoto H, Lively JC, Maeshima Y, Yang C, et al. Physiological levels of tumstatin, a fragment of collagen IV alpha3 chain, are generated by MMP-9 proteolysis and suppress angiogenesis via alphaV beta3 integrin. *Cancer Cell.* (2003) 3:589–601. doi: 10.1016/S1535-6108(03)00133-8
117. Sounni NE, Paye A, Host L, Noël A. MT-MMPS as regulators of vessel stability associated with angiogenesis. *Front Pharmacol.* (2011) 2:111. doi: 10.3389/fphar.2011.00111
118. Lee H, Chang KW, Yang HY, Lin PW, Chen SU, Huang YL. MT1-MMP regulates MMP-2 expression and angiogenesis-related functions in human umbilical vein endothelial cells. *Biochem Biophys Res Commun.* (2013) 437:232–8. doi: 10.1016/j.bbrc.2013.06.046
119. Amălinei C, Căruntu ID, Giuscă SE, Bălan RA. Matrix metalloproteinases involvement in pathologic conditions. *Rom J Morphol Embryol.* (2010) 51:215–28. Available online at: <http://www.rjme.ro/RJME/resources/files/510210215228.pdf>
120. Iizuka S, Ishimaru N, Kudo Y. Matrix metalloproteinases: the gene expression signatures of head and neck cancer progression. *Cancers.* (2014) 6:396–415. doi: 10.3390/cancers6010396
121. Said A, Raufman J-P, Xie G. The role of matrix metalloproteinases in colorectal cancer. *Cancers.* (2014) 6:366–75. doi: 10.3390/cancers6010366
122. Juncker-Jensen A, Deryugina EI, Rimann I, Zajac E, Kupriyanova TA, Engelholm LH, et al. Tumor MMP-1 activates endothelial PAR1 to facilitate

- vascular intravasation and metastatic dissemination. *Cancer Res.* (2013) 73:4196–211. doi: 10.1158/0008-5472.CAN-12-4495
123. Shin DH, Dier U, Melendez JA, Hempel N. Regulation of MMP-1 expression in response to hypoxia is dependent on the intracellular redox status of metastatic bladder cancer cells. *Biochim Biophys Acta - Mol Basis Dis.* (2015) 1852:2593–602. doi: 10.1016/j.bbdis.2015.09.001
 124. Jawad MU, Garamszegi N, Garamszegi SP, Correa-Medina M, Diez JA, Wen R, et al. Matrix metalloproteinase 1: role in sarcoma biology. *PLoS ONE.* (2010) 5:e14250. doi: 10.1371/journal.pone.0014250
 125. Luca M, Huang S, Gershenwald JE, Singh RK, Reich R, Bar-Eli M. Expression of interleukin-8 by human melanoma cells up-regulates MMP-2 activity and increases tumor growth and metastasis. *Am J Pathol.* (1997) 151:1105–13.
 126. Elkin M, Ariel I, Miao HQ, Nagler A, Pines M, De-Groot N, et al. Inhibition of bladder carcinoma angiogenesis, stromal support, and tumor growth by halofuginone. *Cancer Res.* (1999) 59:4111–8.
 127. Partyka R, Gonciarz M, Jałowicki P, Kokocinska D, Byrczek T. VEGF and metalloproteinase 2 (MMP 2) expression in gastric cancer tissue. *Med Sci Monit.* (2012) 18:BR130–4. doi: 10.12659/MSM.882614
 128. Bergers G, Brekken R, McMahon G, Vu TH, Itoh T, Tamaki K, et al. Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat Cell Biol.* (2000) 2:737–44. doi: 10.1038/35036374
 129. Bhoopathi P, Chetty C, Gujrati M, Dinh DH, Rao JS, Lakka SS. The role of MMP-9 in the anti-angiogenic effect of secreted protein acidic and rich in cysteine. *Br J Cancer.* (2010) 102:530–40. doi: 10.1038/sj.bjc.6605538
 130. Takafuji V, Forgues M, Unsworth E, Goldsmith P, Wang XW. An osteopontin fragment is essential for tumor cell invasion in hepatocellular carcinoma. *Oncogene.* (2007) 26:6361–71. doi: 10.1038/sj.onc.1210463
 131. Gupta A, Zhou C, Chellaiyah M. Osteopontin and MMP9: associations with VEGF expression/secretion and angiogenesis in PC3 prostate cancer cells. *Cancers.* (2013) 5:617–38. doi: 10.3390/cancers5020617
 132. Masson V, de la Ballina LR, Munaut C, Wielockx B, Jost M, Maillard C, et al. Contribution of host MMP-2 and MMP-9 to promote tumor vascularization and invasion of malignant keratinocytes. *FASEB J.* (2005) 19:234–6. doi: 10.1096/fj.04.2140fje
 133. Yu Q, Stamenkovic I. Cell surface-localized matrix metalloproteinase-9 proteolytically activates TGF- β and promotes tumor invasion and angiogenesis. *Genes Dev.* (2000) 14:163–76. doi: 10.1101/gad.14.2.163
 134. Webb AH, Gao BT, Goldsmith ZK, Irvine AS, Saleh N, Lee RP, et al. Inhibition of MMP-2 and MMP-9 decreases cellular migration, and angiogenesis in *in vitro* models of retinoblastoma. *BMC Cancer.* (2017) 17:434. doi: 10.1186/s12885-017-3418-y
 135. Agnihotri R, Crawford HC, Haro H, Matrisian LM, Havrda MC, Liaw L. Osteopontin, a novel substrate for matrix metalloproteinase-3 (Stromelysin-1) and matrix metalloproteinase-7 (Matrilysin). *J Biol Chem.* (2001) 276:28261–67. doi: 10.1074/jbc.M103608200
 136. Jin X, Yagi M, Akiyama N, Hirosaki T, Higashi S, Lin C-Y, et al. Matrilysin activates stromelysin (MMP-3) and promotes tumor growth and angiogenesis. *Cancer Sci.* (2006) 97:1327–34. doi: 10.1111/j.1349-7006.2006.00328.x
 137. Kudo Y, Iizuka S, Yoshida M, Tsunematsu T, Kondo T, Subarnhesaj A, et al. Matrix Metalloproteinase-13 (MMP-13) directly and indirectly promotes tumor angiogenesis. *J Biol Chem.* (2012) 287:38716–28. doi: 10.1074/jbc.M112.373159
 138. Patterson BC, Sang QA. Angiostatin-converting enzyme activities of human matrilysin (MMP-7) and gelatinase B/Type IV collagenase (MMP-9). *J Biol Chem.* (1997) 272:28823–5. doi: 10.1074/jbc.272.46.28823
 139. Rorive S, Berton A, D'haene N, Takacs CN, Debeir O, Decaestecker C, et al. Matrix metalloproteinase-9 interplays with the IGFBP2-IGFII complex to promote cell growth and motility in astrocytomas. *Glia.* (2008) 56:1679–90. doi: 10.1002/glia.20719
 140. Azar WJ, Azar SHX, Higgins S, Hu J-F, Hoffman AR, Newgreen DF, et al. IGFBP-2 enhances VEGF gene promoter activity and consequent promotion of angiogenesis by neuroblastoma cells. *Endocrinology.* (2011) 152:3332–42. doi: 10.1210/en.2011-1121
 141. Chan KC, Ko JMY, Lung HL, Sedlacek R, Zhang Z-F, Luo D-Z, et al. Catalytic activity of matrix metalloproteinase-19 is essential for tumor suppressor and anti-angiogenic activities in nasopharyngeal carcinoma. *Int J Cancer.* (2011) 129:1826–37. doi: 10.1002/ijc.25855
 142. Sounni NE, Devy L, Hajitou A, Frankenne F, Munaut C, Gilles C, et al. MT1-MMP expression promotes tumor growth and angiogenesis through an up-regulation of vascular endothelial growth factor expression. *FASEB J.* (2002) 16:555–64. doi: 10.1096/fj.01-0790com
 143. Eisenach PA, Roghi C, Fogarasi M, Murphy G, English WR. MT1-MMP regulates VEGF-A expression through a complex with VEGFR-2 and Src. *J Cell Sci.* (2010) 123:4182–93. doi: 10.1242/jcs.062711
 144. Deryugina EI, Soroceanu L, Strongin AY. Up-regulation of vascular endothelial growth factor by membrane-type 1 matrix metalloproteinase stimulates human glioma xenograft growth and angiogenesis. *Cancer Res.* (2002) 62:580–8. Available online at: <https://cancerres.aacrjournals.org/content/62/2/580.full-text.pdf>
 145. Xu Y, Yuan F-E, Chen Q-X, Liu B-H. Molecular mechanisms involved in angiogenesis and potential target of antiangiogenesis in human glioblastomas. *Glioma.* (2018) 1:35. doi: 10.4103/glioma.glioma_10_17
 146. Basile JR, Holmbeck K, Bugge TH, Gutkind JS. MT1-MMP controls tumor-induced angiogenesis through the release of semaphorin 4D. *J Biol Chem.* (2007) 282:6899–905. doi: 10.1074/jbc.M609570200
 147. Hendrix MJC, Seftor EA, Hess AR, Seftor REB. Vasculogenic mimicry and tumour-cell plasticity: lessons from melanoma. *Nat Rev Cancer.* (2003) 3:411–21. doi: 10.1038/nrc1092
 148. Hawinkels LJAC, Kuiper P, Wiercinska E, Verspaget HW, Liu Z, Pardali E, et al. Matrix Metalloproteinase-14 (MT1-MMP)-mediated endoglin shedding inhibits tumor angiogenesis. *Cancer Res.* (2010) 70:4141–50. doi: 10.1158/0008-5472.CAN-09-4466
 149. Ota I, Li X-Y, Hu Y, Weiss SJ. Induction of a MT1-MMP and MT2-MMP-dependent basement membrane transmigration program in cancer cells by Snail1. *Proc Natl Acad Sci USA.* (2009) 106:20318–23. doi: 10.1073/pnas.0910962106
 150. Liu Y, Sun X, Feng J, Deng L-L, Liu Y, Li B, et al. MT2-MMP induces proteolysis and leads to EMT in carcinomas. *Oncotarget.* (2016) 7:48193–205. doi: 10.18632/oncotarget.10194
 151. Zhao H, Bernardo MM, Osenkowski P, Sohail A, Pei D, Nagase H, et al. Differential Inhibition of Membrane Type 3 (MT3)-Matrix Metalloproteinase (MMP) and MT1-MMP by Tissue Inhibitor of Metalloproteinase (TIMP)-2 and TIMP-3 Regulates Pro-MMP-2 activation. *J Biol Chem.* (2004) 279:8592–601. doi: 10.1074/jbc.M308708200
 152. Chabottaux V, Ricaud S, Host L, Blacher S, Paye A, Thiry M, et al. Membrane-type 4 matrix metalloproteinase (MT4-MMP) induces lung metastasis by alteration of primary breast tumour vascular architecture. *J Cell Mol Med.* (2009) 13:4002–13. doi: 10.1111/j.1582-4934.2009.00764.x
 153. Host L, Paye A, Detry B, Blacher S, Munaut C, Foidart JM, et al. The proteolytic activity of MT4-MMP is required for its pro-angiogenic and pro-metastatic promoting effects. *Int J Cancer.* (2012) 131:1537–48. doi: 10.1002/ijc.27436
 154. Paye A, Truong A, Yip C, Cimino J, Blacher S, Munaut C, et al. EGFR Activation and signaling in cancer cells are enhanced by the membrane-bound metalloprotease MT4-MMP. *Cancer Res.* (2014) 74:6758–70. doi: 10.1158/0008-5472.CAN-13-2994
 155. Sun Q, Weber CR, Sohail A, Bernardo MM, Toth M, Zhao H, et al. MMP25 (MT6-MMP) is highly expressed in human colon cancer, promotes tumor growth, and exhibits unique biochemical properties. *J Biol Chem.* (2007) 282:21998–2010. doi: 10.1074/jbc.M701737200
 156. Chung AS, Lee J, Ferrara N. Targeting the tumour vasculature: Insights from physiological angiogenesis. *Nat Rev Cancer.* (2010) 10:505–14. doi: 10.1038/nrc2868
 157. Meng W, Hao Y, He C, Li L, Zhu G. Exosome-orchestrated hypoxic tumor microenvironment. *Mol Cancer.* (2019) 18:1–14. doi: 10.1186/s12943-019-0982-6
 158. Wang M, Zhao J, Zhang L, Wei F, Lian Y, Wu Y, et al. Role of tumor microenvironment in tumorigenesis. *J Cancer.* (2017) 8:761–73. doi: 10.7150/jca.17648
 159. Blank S, Nienhüser H, Dreikhausen L, Sisic L, Heger U, Ott K, et al. Inflammatory cytokines are associated with response and prognosis in patients with esophageal cancer. *Oncotarget.* (2017) 8:47518–32. doi: 10.18632/oncotarget.17671
 160. Wu X, Tao P, Zhou Q, Li J, Yu Z, Wang X, et al. IL-6 secreted by cancer-associated fibroblasts promotes epithelial-mesenchymal

- transition and metastasis of gastric cancer via JAK2/STAT3 signaling pathway. *Oncotarget*. (2017) 8:20741–50. doi: 10.18632/oncotarget.15119
161. Matsuo Y, Sawai H, Ochi N, Yasuda A, Takahashi H, Funahashi H, et al. Interleukin-1 α secreted by pancreatic cancer cells promotes angiogenesis and its therapeutic implications. *J Surg Res*. (2009) 153:274–81. doi: 10.1016/j.jss.2008.04.040
 162. Huang C, Li Z, Li N, Li Y, Chang A, Zhao T, et al. Interleukin 35 expression correlates with microvessel density in pancreatic ductal adenocarcinoma, recruits monocytes, and promotes growth and angiogenesis of xenograft tumors in mice. *Gastroenterology*. (2018) 154:675–88. doi: 10.1053/j.gastro.2017.09.039
 163. Hosono M, Koma Y-I, Takase N, Urakawa N, Higashino N, Suemune K, et al. CXCL8 derived from tumor-associated macrophages and esophageal squamous cell carcinomas contributes to tumor progression by promoting migration and invasion of cancer cells. *Oncotarget*. (2017) 8:106071–88. doi: 10.18632/oncotarget.22526
 164. Aikins AR, Kim MJ, Raymundo B, Kim CW. Downregulation of transgelin blocks interleukin-8 utilization and suppresses vasculogenic mimicry in breast cancer cells. *Exp Biol Med*. (2017) 242:573–83. doi: 10.1177/1535370216685435
 165. Andisheh-Tadbir A, Mardani M, Pourshahidi S, Nezarati K, Bahadori P. Prognostic value of matrix metalloproteinase-9 expression in oral squamous cell carcinoma and its association with angiogenesis. *J Clin Exp Dent*. (2016) 8:e130–5. doi: 10.4317/jced.52712
 166. Eiró N, Fernandez-García B, González LO, Vizoso FJ. Cytokines related to MMP-11 expression by inflammatory cells and breast cancer metastasis. *Oncimmunology*. (2013) 2:e24010. doi: 10.4161/onci.24010
 167. Lin SC, Hsiao KY, Chang N, Hou PC, Tsai SJ. Loss of dual-specificity phosphatase-2 promotes angiogenesis and metastasis via up-regulation of interleukin-8 in colon cancer. *J Pathol*. (2017) 241:638–48. doi: 10.1002/path.4868
 168. Huang L, Hu B, Ni J, Wu J, Jiang W, Chen C, et al. Transcriptional repression of SOCS3 mediated by IL-6/STAT3 signaling via DNMT1 promotes pancreatic cancer growth and metastasis. *J Exp Clin Cancer Res*. (2016) 35:1–15. doi: 10.1186/s13046-016-0301-7
 169. Shi J, Wei PK. Interleukin-8: a potent promoter of angiogenesis in gastric cancer. *Oncol Lett*. (2016) 11:1043–50. doi: 10.3892/ol.2015.4035
 170. Ruivo CF, Adem B, Silva M, Melo SA. The biology of cancer exosomes: insights and new perspectives. *Cancer Res*. (2017) 77:6480–8. doi: 10.1158/0008-5472.CAN-17-0994
 171. Atrekhany KSN, Drutskaya MS, Nedospasov SA, Grivennikov SI, Kuprash D V. Chemokines, cytokines and exosomes help tumors to shape inflammatory microenvironment. *Pharmacol Ther*. (2016) 168:98–112. doi: 10.1016/j.pharmthera.2016.09.011
 172. Wysoczynski M, Ratajczak MZ. Lung cancer secreted microvesicles: Underappreciated modulators of microenvironment in expanding tumors. *Int J Cancer*. (2009) 125:1595–603. doi: 10.1002/ijc.24479
 173. Sun K, Liao Q, Chen Z, Chen T, Zhang J. Expression of Livin and PlGF in human osteosarcoma is associated with tumor progression and clinical outcome. *Oncol Lett*. (2018) 16:4953–60. doi: 10.3892/ol.2018.9239
 174. Gai C, Carpanetto A, Deregibus MC, Camussi G. Extracellular vesicle-mediated modulation of angiogenesis. *Histol Histopathol*. (2016) 31:379–91. doi: 10.14670/HH-11-708
 175. Loges S, Schmidt T, Carmeliet P. “Antimyoangiogenic” therapy for cancer by inhibiting PlGF. *Clin Cancer Res*. (2009) 15:3648–53. doi: 10.1158/1078-0432.CCR-08-2276
 176. Skog J, Würdinger T, van Rijn S, Meijer DH, Gainche L, Curry WT, et al. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nat Cell Biol*. (2008) 10:1470–76. doi: 10.1038/ncb1800
 177. Klinker MW, Lizzio V, Reed TJ, Fox DA, Lundy SK. Human B cell-derived lymphoblastoid cell lines constitutively produce fas ligand and secrete MHCII(+)/FasL(+) killer exosomes. *Front Immunol*. (2014) 5:144. doi: 10.3389/fimmu.2014.00144
 178. Presta M, Dell’Era P, Mitola S, Moroni E, Ronca R, Rusnati M. Fibroblast growth factor/fibroblast growth factor receptor system in angiogenesis. *Cytokine Growth Factor Rev*. (2005) 16:159–78. doi: 10.1016/j.cytogfr.2005.01.004
 179. Saby C, Collin G, Sinane M, Buache E, Van Gulick L, Saltel F, et al. DDR1 and MT1-MMP expression levels are determinant for triggering BIK-mediated apoptosis by 3D Type I collagen matrix in invasive basal-like breast carcinoma cells. *Front Pharmacol*. (2019) 10:462. doi: 10.3389/fphar.2019.00462
 180. Assent D, Bourgot I, Henny B, Geurts P, Noël A, Foidart JM, et al. A Membrane-type-1 matrix metalloproteinase (MT1-MMP) - Discoidin domain receptor 1 axis regulates collagen-induced apoptosis in breast cancer cells. *PLoS ONE*. (2015) 10:e116006. doi: 10.1371/journal.pone.0116006
 181. Vannitambay A, Seow HJ, Anderson G, Vlahos R, Thompson M, Steinfurt D, et al. Tumour-associated neutrophils and loss of epithelial PTEN can promote corticosteroid-insensitive MMP-9 expression in the chronically inflamed lung microenvironment. *Thorax*. (2017) 72:1140–3. doi: 10.1136/thoraxjnl-2016-209389
 182. Vinnakota K, Zhang Y, Selvanesan BC, Topi G, Salim T, Sand-Dejmek J, et al. M2-like macrophages induce colon cancer cell invasion via matrix metalloproteinases. *J Cell Physiol*. (2017) 232:3468–80. doi: 10.1002/jcp.25808
 183. Bekes EM, Schweighofer B, Kupriyanova TA, Zajac E, Ardi VC, Quigley JP, et al. Tumor-recruited neutrophils and neutrophil TIMP-free MMP-9 regulate coordinately the levels of tumor angiogenesis and efficiency of malignant cell intravasation. *Am J Pathol*. (2011) 179:1455–70. doi: 10.1016/j.ajpath.2011.05.031
 184. De Palma M, Biziato D, Petrova TV. Microenvironmental regulation of tumour angiogenesis. *Nat Rev Cancer*. (2017) 17:457–74. doi: 10.1038/nrc.2017.51
 185. Heusinkveld M, van der Burg SH. Identification and manipulation of tumor associated macrophages in human cancers. *J Transl Med*. (2011) 9:216. doi: 10.1186/1479-5876-9-216
 186. Murdoch C, Muthana M, Coffelt SB, Lewis CE. The role of myeloid cells in the promotion of tumour angiogenesis. *Nat Rev Cancer*. (2008) 8:618–31. doi: 10.1038/nrc2444
 187. Hurt B, Schulick R, Edil B, El Kasmi KC, Barnett C. Cancer-promoting mechanisms of tumor-associated neutrophils. *Am J Surg*. (2017) 214:938–44. doi: 10.1016/j.amjsurg.2017.08.003
 188. Jackute J, Zemaitis M, Pranys D, Sitkauskienė B, Miliauskas S, Vaitkienė S, et al. Distribution of M1 and M2 macrophages in tumor islets and stroma in relation to prognosis of non-small cell lung cancer. *BMC Immunol*. (2018) 19:3. doi: 10.1186/s12865-018-0241-4
 189. Niino D, Komohara Y, Murayama T, Aoki R, Kimura Y, Hashikawa K, et al. Ratio of M2 macrophage expression is closely associated with poor prognosis for Angioimmunoblastic T-cell lymphoma (AITL). *Pathol Int*. (2010) 60:278–83. doi: 10.1111/j.1440-1827.2010.02514.x
 190. Costanza B, Umelo I, Bellier J, Castronovo V, Turtoi A. Stromal modulators of TGF- β in cancer. *J Clin Med*. (2017) 6:7. doi: 10.3390/jcm6010007
 191. Krstic J, Santibanez JF. Transforming growth factor-beta and matrix metalloproteinases: functional interactions in tumor stroma-infiltrating myeloid cells. *Sci World J*. (2014) 2014:521754. doi: 10.1155/2014/521754
 192. Dallas SL, Rosser JL, Mundy GR, Bonewald LF. Proteolysis of latent Transforming Growth Factor- β (TGF- β)-binding Protein-1 by osteoclasts. *J Biol Chem*. (2002) 277:21352–60. doi: 10.1074/jbc.M111663200
 193. Xiong W, Meisinger T, Knispel R, Worth JM, Baxter BT. MMP-2 Regulates Erk1/2 phosphorylation and aortic dilatation in marfan syndrome. *Circ Res*. (2012) 110:e92–101. doi: 10.1161/CIRCRESAHA.112.268268
 194. Lyons RM, Keski-Oja J, Moses HL. Proteolytic activation of latent transforming growth factor- β from fibroblast-conditioned medium. *J Cell Biol*. (1988) 106:1659–65. doi: 10.1083/jcb.106.5.1659
 195. Van den Steen PE, Proost P, Wuyts A, Van Damme J, Opdenakker G. Neutrophil gelatinase B potentiates interleukin-8 tenfold by aminoterminal processing, whereas it degrades CTAP-III, PF-4, and GRO- α and leaves RANTES and MCP-2 intact. *Blood*. (2000) 96:2673–81. doi: 10.1182/blood.V96.8.2673
 196. Xu S, Webb SE, Lau TCK, Cheng SH. Matrix metalloproteinases (MMPs) mediate leukocyte recruitment during the inflammatory phase of zebrafish

- heart regeneration. *Sci Rep.* (2018) 8:1–14. doi: 10.1038/s41598-018-25490-w
197. Lopez T, Mustafa Z, Chen C, Lee KB, Ramirez A, Benitez C, et al. Functional selection of protease inhibitory antibodies. *Proc Natl Acad Sci USA.* (2019) 116:16314–9. doi: 10.1073/pnas.1903330116
198. Larsson P, Syed Khaja AS, Semenas J, Wang T, Sarwar M, Dizeyi N, et al. The functional interlink between AR and MMP9/VEGF signaling axis is mediated through PIP5K1 α /pAKT in prostate cancer. *Int J cancer.* (2019). doi: 10.1002/ijc.32607. [Epub ahead of print].
199. Hassan ZK, Daghestani MH. Curcumin effect on MMPs and TIMPs genes in a breast cancer cell line. *Asian Pac J Cancer Prev.* (2012) 13:3259–64. doi: 10.7314/APJCP.2012.13.7.3259
200. Su C-W, Lin C-W, Yang W-E, Yang S-F. TIMP-3 as a therapeutic target for cancer. *Ther Adv Med Oncol.* (2019) 11:175883591986424. doi: 10.1177/1758835919864247
201. Shailender G, Kumari S, Kiranmayi P, Malla RR. Effect of MMP-2 gene silencing on radiation-induced DNA damage in human normal dermal fibroblasts and breast cancer cells. *Genes Environ Off J Japanese Environ Mutagen Soc.* (2019) 41:16. doi: 10.1186/s41021-019-0131-x
202. Sun X-Y, Han X-M, Zhao X-L, Cheng X-M, Zhang Y. MiR-93-5p promotes cervical cancer progression by targeting THBS2/MMPS signal pathway. *Eur Rev Med Pharmacol Sci.* (2019) 23:5113–21. doi: 10.26355/eurrev_201906_18175
203. Gabelloni P, Da Pozzo E, Bendinelli S, Costa B, Nuti E, Casalini F, et al. Inhibition of metalloproteinases derived from tumours: new insights in the treatment of human glioblastoma. *Neuroscience.* (2010) 168:514–22. doi: 10.1016/j.neuroscience.2010.03.064
204. Frantz C, Stewart KM, Weaver VM. The extracellular matrix at a glance. *J Cell Sci.* (2010) 123:4195–200. doi: 10.1242/jcs.023820
205. Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. *J Clin Invest.* (2009) 119:1420–8. doi: 10.1172/JCI39104

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Quintero-Fabián, Arreola, Becerril-Villanueva, Torres-Romero, Arana-Argáez, Lara-Riegos, Ramírez-Camacho and Alvarez-Sánchez. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.