



Fourier transform infrared difference spectroscopy for studying the molecular mechanism of photosynthetic water oxidation

Hsiu-An Chu*

Institute of Plant and Microbial Biology, Academia Sinica, Taipei, Taiwan

Edited by:

Harvey J. M. Hou, Alabama State University, USA

Reviewed by:

Takumi Noguchi, Nagoya University, Japan

Richard Debus, University of California at Riverside, USA

***Correspondence:**

Hsiu-An Chu, Institute of Plant and Microbial Biology, Academia Sinica, Taipei 11529, Taiwan.
e-mail: chuha@gate.sinica.edu.tw

The photosystem II reaction center mediates the light-induced transfer of electrons from water to plastoquinone, with concomitant production of O₂. Water oxidation chemistry occurs in the oxygen-evolving complex (OEC), which consists of an inorganic Mn₄CaO₅ cluster and its surrounding protein matrix. Light-induced Fourier transform infrared (FTIR) difference spectroscopy has been successfully used to study the molecular mechanism of photosynthetic water oxidation. This powerful technique has enabled the characterization of the dynamic structural changes in active water molecules, the Mn₄CaO₅ cluster, and its surrounding protein matrix during the catalytic cycle. This mini-review presents an overview of recent important progress in FTIR studies of the OEC and implications for revealing the molecular mechanism of photosynthetic water oxidation.

Keywords: photosystem II, oxygen evolution, FTIR, manganese cluster, infrared spectroscopy

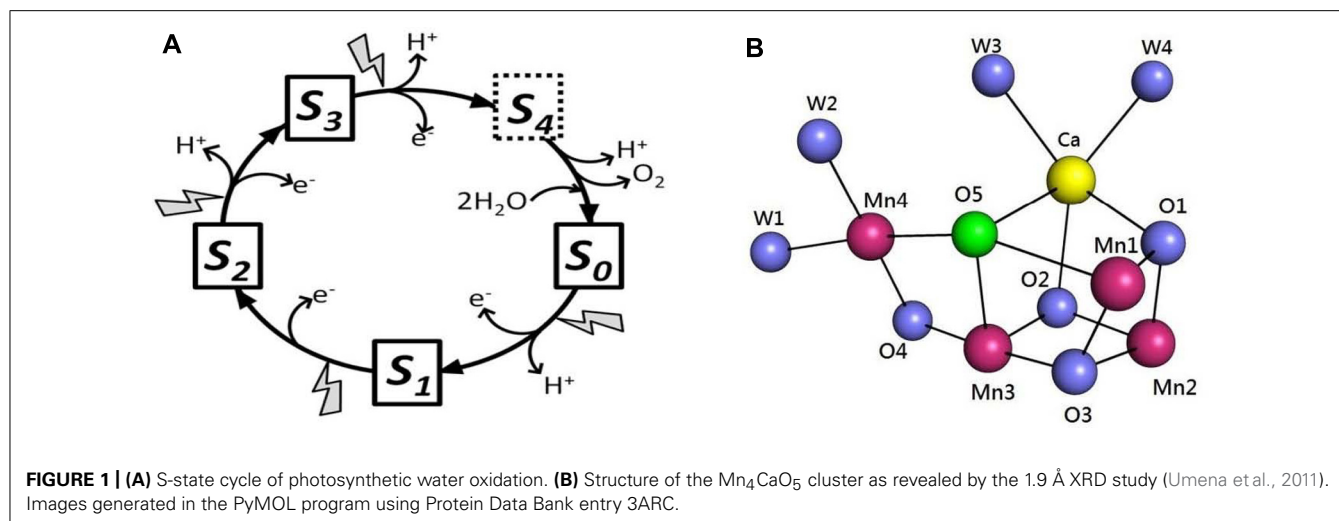
INTRODUCTION

Photosynthetic water oxidation is catalyzed by a Mn₄Ca cluster and its surrounding protein matrix in photosystem II (PSII; Ferreira et al., 2004; Loll et al., 2005; Yano et al., 2006; Umena et al., 2011). The oxygen-evolving complex (OEC) accumulates oxidizing equivalents from the photochemical reactions within PSII and cycles through five oxidation states, termed S_n (n = 0–4, n representing the storage of oxidizing equivalents in the OEC; Kok et al., 1970; Figure 1A). The molecule O₂ is produced in the transition of S₃-(S₄)-S₀. One of the recent major breakthroughs in PSII research was the report of the crystal structure of oxygen-evolving PSII at 1.9 Å resolution (Umena et al., 2011). The structure of the Mn₄CaO₅ cluster is shown in Figure 1B. Three Mn, one Ca, and four oxygen atoms form a cubane-like structure; the fourth Mn connects to the cubic structure by two μ-oxo-bridges. The Mn₄CaO₅ cluster is connected with four water molecules: two are ligated to Ca and two to Mn₄ (Umena et al., 2011; Figure 1B). These water molecules are candidates for substrates in photosynthetic water oxidation. Another distinct feature in the structure is the apparently longer bond distances between the O₅-bridging oxygen atom and neighboring metal ions, which indicates weak bonding of this oxygen atom in the cluster. O₅ was proposed as a candidate for one of the substrates in dioxygen formation (Umena et al., 2011). More recent studies have suggested that the structure of the X-ray diffraction (XRD) model of PSII is modified by radiation-induced reduction of the Mn cluster (Luber et al., 2011; Grundmeier and Dau, 2012). Despite this problem, the 1.9 Å XRD structure is the crucial foundation for spectroscopic and mechanistic studies of photosynthetic water oxidation.

Abbreviations: EPR, electron paramagnetic resonance; FTIR, Fourier transform infrared; OEC, oxygen-evolving complex; OTG, octyl-β-D-thioglucoopyranoside; PSII, photosystem II; Q_A, primary quinone electron acceptor in PSII; XRD, X-ray diffraction.

Several structural models of photosynthetic water oxidation have been proposed (Hoganson and Babcock, 1997; Pecoraro et al., 1998; McEvoy and Brudvig, 2006; Kusunoki, 2007; Pushkar et al., 2008; Siegbahn, 2009; Grundmeier and Dau, 2012), with differing locations and molecular structures of S-state intermediates (i.e., terminal water molecules or water-derived metal ligands). Several molecular spectroscopic techniques, including advanced electron paramagnetic resonance (EPR) spectroscopy (Britt et al., 2004; McConnell et al., 2011; Rapatskiy et al., 2012) and light-induced Fourier transform infrared (FTIR) difference spectroscopy (see below) have been extensively used to resolve this issue.

Fourier transform infrared difference spectroscopy has been widely used to study the structural changes in the OEC during the S-state catalytic cycle. The S₂-minus-S₁ mid-frequency (1800–1000 cm⁻¹) FTIR difference spectrum was first reported in 1992 (Noguchi et al., 1992). The S₃-minus-S₂ spectrum of the PSII/OEC was reported in 2000 (Chu et al., 2000b), and spectra of flash-induced S-state transitions (S₁ → S₂, S₂ → S₃, S₃ → S₀, and S₀ → S₁) during the complete S-state cycle were reported 1 year later (Hillier and Babcock, 2001; Noguchi and Sugiura, 2001). Many FTIR studies of the OEC focused on the mid-frequency region (1800–1000 cm⁻¹) of the IR spectrum, which contains information on structural changes of protein backbones and amino acid side-chains associated with S-state transitions of the OEC. One very important development in FTIR studies of the OEC were reports of high-frequency spectra (3700–3500 cm⁻¹) of the OEC, which contain information on structural changes of the weakly H-bonded OH-stretching of active water molecules during S-state transitions of the OEC (Noguchi and Sugiura, 2000, 2002a,b). The other important developments were reports of low-frequency spectra (<1000 cm⁻¹), which contain information on metal-ligand and manganese-substrate vibration modes



of the OEC (Chu et al., 1999, 2000a,c; Yamanari et al., 2004; Kimura et al., 2005b).

This mini-review gives an overview of recent important progress in FTIR studies of the OEC, combined with new spectroscopic and XRD structural information, to understand the chemical mechanism of photosynthetic water oxidation. More comprehensive reviews on FTIR studies of the OEC are available (Noguchi, 2007, 2008a,b; Debus, 2008).

OH-STRETCHING VIBRATIONAL MODES OF ACTIVE WATER MOLECULES IN THE HIGH-FREQUENCY REGION ($3700\text{--}3500\text{ cm}^{-1}$) OF THE OEC

The reactions of substrate water during the S-state catalytic cycle of the OEC are of paramount importance to understand the chemical mechanism of photosynthetic water oxidation. In the new XRD structure of the Mn_4CaO_5 cluster, the four water molecules connected to the OEC are involved in a hydrogen-bonded network linking the Mn_4CaO_5 -cluster and Y_Z (Umena et al., 2011). The bond distances (2.8–3.3 Å) between oxygen atoms of coordinated water molecules and their neighboring water molecules indicate that most of the O–H groups of the water molecules are weakly hydrogen bonded and will appear in the weakly hydrogen-bonded OH-stretch ($3750\text{--}3500\text{ cm}^{-1}$) region of the FTIR spectra. Noguchi and colleagues reported flash-induced difference spectra of S-state transitions in the weakly H-bonded OH-stretching region (Noguchi and Sugiura, 2000, 2002a,b). One active water molecule on the OEC, which gave rise to the S_1 band at $\sim 3585\text{ cm}^{-1}$ and the S_2 band at $\sim 3618\text{ cm}^{-1}$, was identified at 250 K in light-induced S_2/S_1 FTIR difference spectrum (Noguchi and Sugiura, 2000) and during the S-state cycle at 10°C (Noguchi and Sugiura, 2002a,b) in PSII core complexes from *Thermosynechococcus elongatus*. The results indicated a weakened hydrogen bond of the OH group for one water molecule connected to the OEC during the $S_1 \rightarrow S_2$ transition. In contrast to the $S_1 \rightarrow S_2$ transition, the $S_2 \rightarrow S_3$, $S_3 \rightarrow S_0$, and $S_0 \rightarrow S_1$ transitions all showed a negative OH-stretching mode at different frequencies, which indicates that these water (or hydroxide) molecules were involved in proton release reactions of the OEC or formed strong

hydrogen-bonding interactions during these transitions (Noguchi and Sugiura, 2002a,b). In addition, these observations are consistent with a recent FTIR study which concluded that the proton release pattern from the substrate water on the OEC is in 1:0:1:2 stoichiometry for the $S_0 \rightarrow S_1 \rightarrow S_2 \rightarrow S_3 \rightarrow S_0$ transition (Suzuki et al., 2009). One of the important issues is the exact location of these active water molecules detected by FTIR difference spectra on the OEC (e.g., associated with Mn or Ca) in the new XRD structure.

Mn-LIGAND AND Mn-SUBSTRATE VIBRATION MODES IN THE LOW-FREQUENCY REGION ($<1000\text{ cm}^{-1}$) OF THE OEC

From studies of Mn model compound, Mn-ligand and Mn-substrate vibration modes of the PSII/OEC are expected to show up in the low-frequency region ($<1000\text{ cm}^{-1}$) of the IR spectrum (Chu et al., 2001c). In low-frequency S_2/S_1 FTIR difference spectra of octyl- β -D-thioglucopyranoside (OTG) PSII core preparations of spinach, a positive mode at 606 cm^{-1} in ^{16}O water clearly downshifted to 596 cm^{-1} in ^{18}O water (Chu et al., 2000c; **Figure 2A**). With double-difference (S_2/S_1 and ^{16}O minus ^{18}O) spectra, the 606 cm^{-1} mode was assigned to an S_2 mode, and a corresponding S_1 mode at about 625 cm^{-1} was identified (Chu et al., 2000c). In addition, this 606-cm^{-1} mode was up-shifted to about 618 cm^{-1} with Sr^{2+} substitution but not significantly affected by ^{44}Ca isotope substitution (Chu et al., 2000c; Kimura et al., 2005a; **Figure 2B**). From these results and studies of Mn model compounds, this vibrational mode at 606 cm^{-1} in the S_2 state was assigned to a Mn–O–Mn cluster vibration in the OEC (Chu et al., 2000c). The structure of this Mn–O–Mn cluster very likely includes additional oxo and carboxylate bridges(s). IR modes for $\nu(\text{Mn}=\text{O})$ and $\nu_{\text{asy}}(\text{Mn}-\text{O}-\text{Mn})$ for a singly oxo-bridged Mn cluster usually occur at $>700\text{ cm}^{-1}$ and typically have a $30\text{--}40\text{ cm}^{-1}$ downshift (Chu et al., 2001c). They are unlikely to be the origin of the 606-cm^{-1} mode. Furthermore, this 606-cm^{-1} mode was altered in S_2/S_1 FTIR difference spectra of Ala344D1Gly, Glu189Gln, and Asp170HisD1 *Synechocystis* mutant PSII particles (Chu et al., 2001a; Mizusawa et al., 2004; Kimura et al., 2005c). All the above amino acid residues are direct ligands for the Mn_4Ca

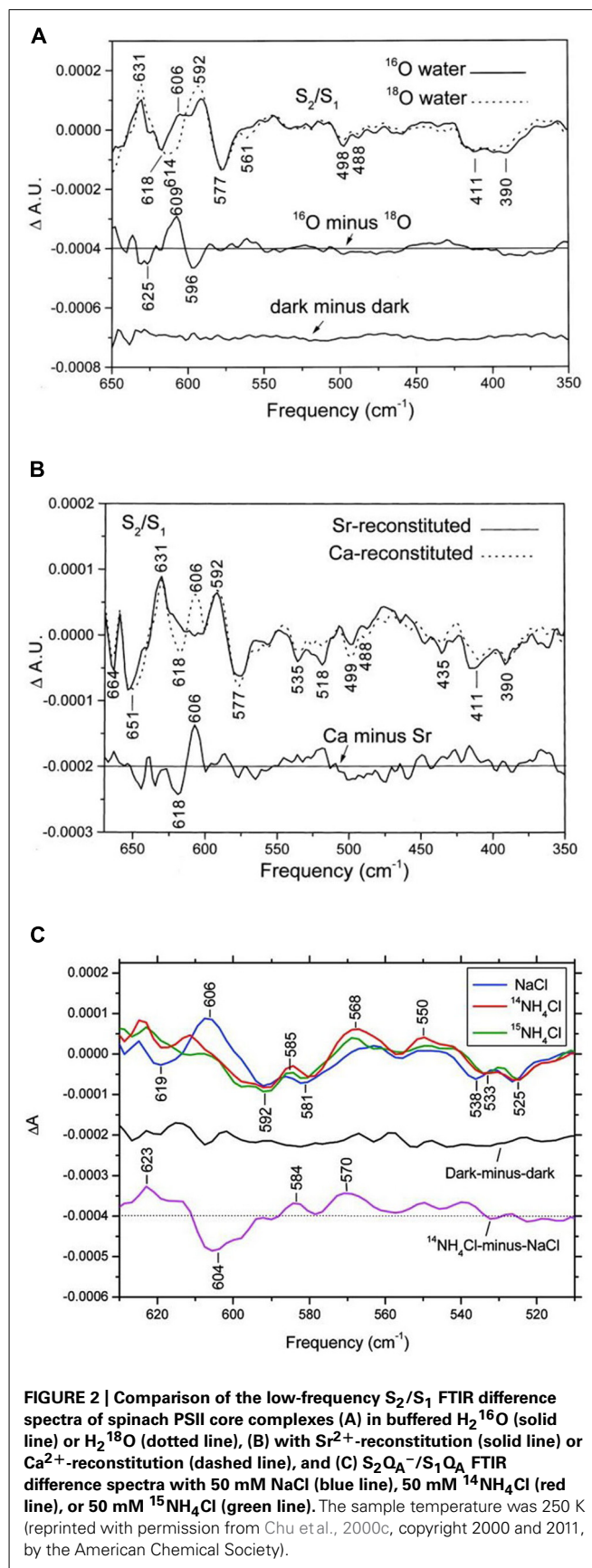
cluster. Therefore, the structure of the Mn–O–Mn cluster is structurally coupled to its surrounding ligand environment.

Low-frequency S_3/S_2 spectra were reported in OTG PSII core preparations of spinach, in which intense bands at 604(–) and 621(+) cm^{-1} were sensitive to ^{18}O water exchange (Chu et al., 2001b). The S_3 mode at $\sim 621 \text{ cm}^{-1}$ was attributed to the Mn–O–Mn cluster mode of the S_3 state. Kimura et al. (2005b) reported on $^{16}\text{O}/^{18}\text{O}$ - and/or H/D water-sensitive low-frequency vibrations of the OEC during the complete S-state cycle in PSII core particles from *T. elongatus*. The S_2 mode at $\sim 606 \text{ cm}^{-1}$ changed their sign and intensity during S-state cycling, which indicates S-state-dependent changes in the core structure of the Mn_4CaO_5 cluster. In addition, several IR bands sensitive to both $^{16}\text{O}/^{18}\text{O}$ and H/D exchanges were attributed to S-state intermediates during the S-state cycling (Kimura et al., 2005b). Furthermore, an intense 577($-$) cm^{-1} band in the S_2/S_1 spectra was found insensitive to universal ^{15}N - and ^{13}C -isotope labeling and assigned to the skeletal vibration of the Mn cluster or stretching vibrational modes of the Mn ligand (Kimura et al., 2003).

Low-frequency FTIR results demonstrate that one bridged oxygen atom in the Mn–O–Mn cluster of the OEC is accessible to and can be exchanged with bulky-phase water. This exchange occurs within minutes or faster because it is complete within 30 min (Chu et al., 2000c). A recent study involving W-band ^{17}O electron–electron double resonance–detected nuclear magnetic resonance (NMR) spectroscopy reported that one μ -oxo bridge of the OEC can exchange with H_2^{17}O on a time scale ($\leq 15 \text{ s}$) similar to that of substrate water on the OEC (Rapatskiy et al., 2012). This study also suggested that the exchangeable μ -oxo bridge links the outer Mn to the $\text{Mn}_3\text{O}_3\text{Ca}$ open-cuboidal unit (O_4 and O_5 in Figure 1). The authors of this study favored the Ca-linked O_5 oxygen assignment (Rapatskiy et al., 2012). Low-frequency FTIR results showed that the Mn–O–Mn cluster mode at 606 cm^{-1} is sensitive to Sr^{2+} substitution but not ^{44}Ca substitution (Chu et al., 2000c; Kimura et al., 2005a). Considering the structure of O_5 in the Mn_4CaO_5 cluster (Umena et al., 2011; Figure 1B), the ^{44}Ca -induced isotopic shift of the Mn–O–Mn cluster mode may have been too small to be detected by previous FTIR studies. Thus, the O_5 -bridging oxygen atom is a good candidate for the exchangeable-bridged oxygen atom in the Mn–O–Mn cluster identified by FTIR. A recent continue-wave Q-band electron nuclear double resonance (ENDOR) study reported a much slower ^{17}O exchange rate (on the time scale of hours) with ^{17}O -labeled water into the μ -oxo bridge of the OEC (McConnell et al., 2011). Future study is required to resolve this discrepancy.

EFFECT OF AMMONIA ON THE OEC

Because of the structural similarity between NH_3 and H_2O and the ability of NH_3 to inhibit photosynthetic water oxidation, the NH_3 binding site on the OEC might occur at the substrate water-binding site. Previous EPR studies of NH_3 -treated PSII samples demonstrated that the S_2 -state multiline EPR signal is altered when samples illuminated at 200 K are subsequently “annealed” above 250 K (Beck et al., 1986; Britt et al., 1989). FTIR studies showed that NH_3 induced characteristic spectral changes in the S_2/S_1 spectra at 250 K (Chu et al., 2004a; Fang et al., 2005). Among them, the S_2 -state symmetric carboxylate stretching mode at 1365 cm^{-1} in



the S_2/S_1 spectrum of control samples up-shifted to $\sim 1379\text{ cm}^{-1}$ in NH_3 -treated samples. This carboxylate mode was also altered by Sr^{2+} substitution (Strickler et al., 2005; Suzuki et al., 2006), which indicates that the action site of NH_3 on the OEC is near the Ca^{2+} site. In addition, the conditions that give rise to the NH_3 -induced up-shift of this S_2 -state carboxylate stretching mode at 1365 cm^{-1} are strongly correlated with those producing the modified S_2 -state multiline EPR signal (Chu et al., 2004a; Fang et al., 2005). Furthermore, a recent FTIR result showed that NH_3 did not replace the active water molecule connected to the OEC during the S_1 -to- S_2 transition at 250 K, whereas the Mn–O–Mn cluster vibrational mode at 606 cm^{-1} was diminished or underwent a large shift (Hou et al., 2011; **Figure 2C**). The above results are consistent with the proposal that NH_3 may replace one of the bridging oxygen atoms, presumably O_5 , in the Mn_4CaO_5 cluster during the S_1 -to- S_2 transition (Britt et al., 1989).

The other intriguing FTIR finding is that the effect of NH_3 -induced up-shift of 1365 cm^{-1} mode in the S_2/S_1 spectrum was diminished at temperatures above 0°C (Huang et al., 2008). The results indicate that the interaction of NH_3 with the OEC is attenuated at temperatures above 0°C (Huang et al., 2008). In addition, a recent FTIR study reported an inhibitory effect of the ammonium cation on the PSII/OEC at 283 K (Tsuno et al., 2011). The results suggested that the ammonium cation perturbs some carboxylate residues coupled to the Mn cluster during the S_1 -to- S_2 transition and inhibits the oxygen evolution reaction at 283 K (Tsuno et al., 2011).

FTIR RESULTS FOR PROTEIN LIGANDS OF THE OEC

Fourier transform infrared studies involving isotopic labeling and site-directed mutagenesis have provided a wealth of information on dynamic structural changes of the protein backbones and amino acid side-chains during the S-state transitions of the OEC (Debus, 2008; Noguchi, 2008a; Shimada et al., 2011). An isotope-edited FTIR study identified the L-[$1\text{-}^{13}\text{C}$]alanine-sensitive symmetric carboxylate stretching modes in S_2/S_1 difference spectra to the $\alpha\text{-COO}^-$ group of D1-Ala344 (Chu et al., 2004b). This mode appears at $\sim 1356\text{ cm}^{-1}$ in the S_1 state and at ~ 1339 or $\sim 1320\text{ cm}^{-1}$ in the S_2 state in unlabeled wild-type PSII particles but not in D1-Ala344Gly and D1-Ala344Ser mutant PSII particles. These frequencies are consistent with unidentate ligation of the $\alpha\text{-COO}^-$ group of D1-Ala344 to the Mn_4Ca cluster in both the S_1 and S_2 state (Chu et al., 2004b; Strickler et al., 2005). In addition, substituting Sr for Ca did not alter the symmetric carboxylate stretching modes of D1-Ala344 (Strickler et al., 2005). The results suggested that the $\alpha\text{-COO}^-$ group of D1-Ala344 did not ligate Ca. In the 1.9 \AA XRD structure, the $\alpha\text{-COO}^-$ group of D1-Ala344 shows very asymmetrical bridging between Mn_2 and Ca in the cluster, with the Mn–O distance 2.0 \AA and Ca–O distance 2.6 \AA (Kawakami et al., 2011). In addition, the isotopic bands for the $\alpha\text{-COO}^-$ group of D1-Ala344 showed characteristic changes during S-state cycling (Kimura et al., 2005d). These results indicated that the C-terminal Ala 344D1 is structurally coupled, presumably directly ligated, to the Mn ion that undergoes oxidation of Mn(III) to Mn(IV) during the S_1 -to- S_2 transition and is reduced in reverse with the S_3 -to- S_0 transition (Chu et al., 2004b; Kimura et al., 2005d). In contrast, mutations of D1-Asp170,

D1-Glu189, and D1-Asp342 did not eliminate any carboxylate vibrational stretching modes during S-state cycling of the OEC (Debus et al., 2005; Strickler et al., 2006, 2007). Recent computational studies suggested that vibrations of carboxylate ligands can be quite insensitive to Mn oxidation, if they are not coordinated along the Jahn–Teller axis (Sproviero et al., 2008). In their model, the only amino acid residue that is ligated along the Jahn–Teller axis of a Mn^{III} ion is CP43-E354.

Of note, CP43-E354Q mutant PSII particles gave rise to characteristic spectral changes in the amide and carboxylate stretch regions of FTIR difference spectra during S-state transitions (Strickler et al., 2008; Shimada et al., 2009; Service et al., 2010). In addition, the weakly H-bonded O–H stretching modes of the active water molecule associated with the OEC were significantly altered in S_2/S_1 FTIR difference spectra of CP43-E354Q mutant PSII particles (Shimada et al., 2009). Furthermore, H_2^{18}O exchange mass spectrometry experiments showed that the CP43-E354Q mutation weakened the binding of both substrate-water molecules (or water-derived ligands), particularly affecting the one with faster exchange in the S_3 state (Service et al., 2010). The XRD structure of the OEC showed that coordinated water molecules were on Ca^{2+} and Mn_4 , which were both not ligated by CP43-E354 (Umena et al., 2011). Presumably, CP43-E354Q mutation may induce significant structural changes to the Mn_4CaO_5 core that affects associated active water molecule(s) on the OEC during the S_1 -to- S_2 transition.

A recent time-resolved infrared study revealed the proton and protein dynamics associated with the OEC during the S-state transitions (Noguchi et al., 2012). The results suggest that during the S_3 -to- S_0 transition, protons are greatly rearranged to form a transient state before the oxidation of the Mn_4CaO_5 cluster that leads to O_2 formation. In addition, an early proton movement was detected during the $S_2 \rightarrow S_3$ transition, indicating a proton release coupled with the electron transfer reaction. Furthermore, a relatively slow carboxylate movement occurred in the $S_0 \rightarrow S_1$ transition, which might reflect the protein relaxation process to stabilize the S_1 state (Noguchi et al., 2012). This study demonstrates that time-resolved infrared technique is extremely useful to monitor proton and protein dynamics of the OEC during photosynthetic oxygen evolution.

BIOINORGANIC MODELS FOR FTIR SPECTRAL INTERPRETATION

Vibrational data from model compounds relevant to the OEC is crucial to interpret FTIR data of the OEC during S-state cycling. However, vibrational data for synthetic multinuclear Mn complexes are still limited (Cua et al., 2003; Berggren et al., 2012). Particularly, vibrational data are needed for the Ca–Mn multinuclear cluster that models the Mn_4CaO_5 cluster (Kanady et al., 2011; Mukherjee et al., 2012). One previous study reported IR spectra and normal mode analysis of the adamantane-like complex $[\text{Mn}_4\text{O}_6(\text{bpea})_4]^{n+}$ (Visser et al., 2002). By using the electrochemical method to record the difference IR spectrum and ^{18}O isotopic labeling, the authors identified Mn–O vibrational modes for $[\text{Mn}^{\text{IV}}_4]$ and $[\text{Mn}^{\text{IV}}_3\text{Mn}^{\text{III}}]$. Comparison with OEC data ruled out the adamantane-like complex as the possible structure

intermediate. Nevertheless, this approach is very powerful for interpreting FTIR data for the OEC during S-state cycling.

CONCLUSIONS AND PERSPECTIVES

Light-induced FTIR difference spectroscopy has become a fruitful structural technique to study the molecular mechanism of photosynthetic water oxidation. The new high-resolution XRD structure of the OEC has served as a crucial foundation for designing FTIR experiments and interpreting FTIR data. Combined with isotopic labeling, site-directed mutagenesis, model

compound studies, and normal mode analysis, FTIR difference spectroscopy will continue to provide important structural and mechanistic insights into the water-splitting process in PSII.

ACKNOWLEDGMENTS

The author is grateful to Prof. Richard J. Debus for helpful suggestions on the manuscript. This work was supported by National Science Council in Taiwan (NSC 101-2627-M-001-001) and by Academia Sinica.

REFERENCES

- Beck, W. F., De Paula, J. C., and Brudvig, G. W. (1986). Ammonia binds to the manganese site of the oxygen-evolving complex of photosystem II in the S2 state. *J. Am. Chem. Soc.* 108, 4018–4022.
- Berggren, G., Anderlund, M. F., Styring, S., and Thapper, A. (2012). FTIR study of manganese dimers with carboxylate donors as model complexes for the water oxidation complex in photosystem II. *Inorg. Chem.* 51, 2332–2337.
- Britt, R. D., Campbell, K. A., Peloquin, J. M., Gilchrist, M. L., Aznar, C. P., Dicus, M. M., et al. (2004). Recent pulsed EPR studies of the photosystem II oxygen-evolving complex: implications as to water oxidation mechanisms. *Biochim. Biophys. Acta* 1655, 158–171.
- Britt, R. D., Zimmermann, J. L., Sauer, K., and Klein, M. P. (1989). Ammonia binds to the catalytic manganese of the oxygen-evolving complex of photosystem II. Evidence by electron spin-echo envelope modulation spectroscopy. *J. Am. Chem. Soc.* 111, 3522–3532.
- Chu, H.-A., Debus, R. J., and Babcock, G. T. (2001a). D1-Asp170 is structurally coupled to the oxygen evolving complex in photosystem II as revealed by light-induced Fourier transform infrared difference spectroscopy. *Biochemistry* 40, 2312–2316.
- Chu, H.-A., Hillier, W., Law, N. A., and Babcock, G. T. (2001b). "Identification of a possible Mn–O–Mn cluster vibrational mode of the S3 state in the oxygen-evolving complex of photosystem II by the low-frequency FTIR spectroscopy," in *PS2001 proceedings: 12th International Congress on Photosynthesis, S13-027*, CSIRO Publishing, Collingwood, Australia.
- Chu, H.-A., Hillier, W., Law, N. A., and Babcock, G. T. (2001c). Vibrational spectroscopy of the oxygen-evolving complex and of manganese model compounds. *Biochim. Biophys. Acta* 1503, 69–82.
- Chu, H.-A., Feng, Y.-W., Wang, C.-M., Chiang, K.-A., and Ke, S.-C. (2004a). Ammonia-induced structural changes of the oxygen-evolving complex in photosystem II as revealed by light-induced FTIR difference spectroscopy. *Biochemistry* 43, 10877–10885.
- Chu, H.-A., Hillier, W., and Debus, R. J. (2004b). Evidence that the C-terminus of the D1 polypeptide of photosystem II is ligated to the manganese ion that undergoes oxidation during the S1 to S2 transition: an isotope-edited FTIR study. *Biochemistry* 43, 3152–3166.
- Chu, H.-A., Gardner, M., Hillier, W., and Babcock, G. (2000a). Low-frequency Fourier transform infrared spectroscopy of the oxygen-evolving complex in photosystem II. *Photosynth. Res.* 66, 57–63.
- Chu, H.-A., Hillier, W., Law, N. A., Sackett, H., Haymond, S., and Babcock, G. T. (2000b). Light-induced FTIR difference spectroscopy of the S2-to-S3 state transition of the oxygen-evolving complex in photosystem II. *Biochim. Biophys. Acta* 1459, 528–532.
- Chu, H.-A., Sackett, H., and Babcock, G. T. (2000c). Identification of a Mn–O–Mn cluster vibrational mode of the oxygen-evolving complex in photosystem II by low-frequency FTIR spectroscopy. *Biochemistry* 39, 14371–14376.
- Chu, H.-A., Gardner, M. T., O'Brien, J. P., and Babcock, G. T. (1999). Low-frequency Fourier transform infrared spectroscopy of the oxygen-evolving and quinone acceptor complexes in photosystem II. *Biochemistry* 38, 4533–4541.
- Cua, A., Vrettos, J., Paula, J., Brudvig, G., and Bocian, D. (2003). Raman spectra and normal coordinate analyses of low-frequency vibrations of oxo-bridged manganese complexes. *J. Biol. Inorg. Chem.* 8, 439–451.
- Debus, R. J. (2008). Protein ligation of the photosynthetic oxygen-evolving center. *Coord. Chem. Rev.* 252, 244–258.
- Debus, R. J., Strickler, M. A., Walker, L. M., and Hillier, W. (2005). No evidence from FTIR difference spectroscopy that aspartate-170 of the D1 polypeptide ligates a manganese ion that undergoes oxidation during the S0 to S1, S1 to S2, or S2 to S3 transitions in photosystem II. *Biochemistry* 44, 1367–1374.
- Fang, C.-H., Chiang, K.-A., Hung, C.-H., Chang, K., Ke, S.-C., and Chu, H.-A. (2005). Effects of ethylene glycol and methanol on ammonia-induced structural changes of the oxygen-evolving complex in photosystem II. *Biochemistry* 44, 9758–9765.
- Ferreira, K. N., Iverson, T. M., Maghlaoui, K., Barber, J., and Iwata, S. (2004). Architecture of the photosynthetic oxygen-evolving center. *Science* 303, 1831–1838.
- Grundmeier, A., and Dau, H. (2012). Structural models of the manganese complex of photosystem II and mechanistic implications. *Biochim. Biophys. Acta* 1817, 88–105.
- Hillier, W., and Babcock, G. T. (2001). S-state dependent Fourier transform infrared difference spectra for the photosystem II oxygen evolving complex. *Biochemistry* 40, 1503–1509.
- Hoganson, C. W., and Babcock, G. T. (1997). A metalloradical mechanism for the generation of oxygen from water in photosynthesis. *Science* 277, 1953–1956.
- Hou, L.-H., Wu, C.-M., Huang, H.-H., and Chu, H.-A. (2011). Effects of ammonia on the structure of the oxygen-evolving complex in photosystem II as revealed by light-induced FTIR difference spectroscopy. *Biochemistry* 50, 9248–9254.
- Huang, H.-H., Wang, T.-H., and Chu, H.-A. (2008). "Ammonia-induced structural changes of the oxygen-evolving complex in photosystem II diminished at 277 K as revealed by light-induced FTIR difference Spectroscopy," in *Photosynthesis. Energy from the Sun*, eds J. Allen, E. Gantt, J. Golbeck, and B. Osmond (Dordrecht: Springer), 389–391.
- Kanady, J. S., Tsui, E. Y., Day, M. W., and Agapie, T. (2011). A synthetic model of the Mn₃Ca subsite of the oxygen-evolving complex in photosystem II. *Science* 333, 733–736.
- Kawakami, K., Umena, Y., Kamiya, N., and Shen, J.-R. (2011). Structure of the catalytic, inorganic core of oxygen-evolving photosystem II at 1.9 Å resolution. *J. Photochem. Photobiol. B Biol.* 104, 9–18.
- Kimura, Y., Hasegawa, K., Yamanari, T., and Ono, T.-A. (2005a). Studies on photosynthetic oxygen-evolving complex by means of Fourier transform infrared spectroscopy: calcium and chloride cofactors. *Photosynth. Res.* 84, 245–250.
- Kimura, Y., Ishii, A., Yamanari, T., and Ono, T.-A. (2005b). Water-sensitive Low-frequency vibrations of reaction intermediates during S-state cycling in photosynthetic water oxidation. *Biochemistry* 44, 7613–7622.
- Kimura, Y., Mizusawa, N., Ishii, A., Nakazawa, S., and Ono, T. A. (2005c). Changes in structural and functional properties of oxygen-evolving complex induced by replacement of D1-glutamate 189 with glutamine in photosystem II: ligation of glutamate 189 carboxylate to the manganese cluster. *J. Biol. Chem.* 280, 37895–37900.
- Kimura, Y., Mizusawa, N., Yamanari, T., Ishii, A., and Ono, T. A. (2005d). Structural changes of D1 C-terminal alpha-carboxylate during S-state cycling in photosynthetic oxygen evolution. *J. Biol. Chem.* 280, 2078–2083.
- Kimura, Y., Mizusawa, N., Ishii, A., Yamanari, T., and Ono, T.-A. (2003). Changes of low-frequency vibrational modes induced by universal ¹⁵N- and ¹³C-isotope labeling in S2/S1 FTIR difference spectrum of oxygen-evolving complex. *Biochemistry* 42, 13170–13177.
- Kok, B., Forbush, B., and McGloin, M. (1970). Cooperation of charges in photosynthetic O₂ evolution-I. A linear four step mechanism. *Photochem. Photobiol.* 11, 457–475.

- Kusunoki, M. (2007). Mono-manganese mechanism of the photosystem II water splitting reaction by a unique Mn_4Ca cluster. *Biochim. Biophys. Acta* 1767, 484–492.
- Loll, B., Kern, J., Saenger, W., Zouni, A., and Biesiadka, J. (2005). Towards complete cofactor arrangement in the 3.0 Å resolution structure of photosystem II. *Nature* 438, 1040–1044.
- Luber, S., Rivalta, I., Umena, Y., Kawakami, K., Shen, J.-R., Kamiya, N., et al. (2011). S1-state model of the O_2 -evolving complex of photosystem II. *Biochemistry* 50, 6308–6311.
- McConnell, I. L., Grigoryants, V. M., Scholes, C. P., Myers, W. K., Chen, P.-Y., Whittaker, J. W., et al. (2011). EPR-ENDOR characterization of (^{17}O , 1H , 2H) water in manganese catalase and its relevance to the oxygen-evolving complex of photosystem II. *J. Am. Chem. Soc.* 134, 1504–1512.
- McEvoy, J. P., and Brudvig, G. W. (2006). Water-splitting chemistry of photosystem II. *Chem. Rev.* 106, 4455–4483.
- Mizusawa, N., Kimura, Y., Ishii, A., Yamanari, T., Nakazawa, S., Teramoto, H., et al. (2004). Impact of replacement of D1 C-terminal alanine with glycine on structure and function of photosynthetic oxygen-evolving complex. *J. Biol. Chem.* 279, 29622–29627.
- Mukherjee, S., Stull, J. A., Yano, J., Stamatatos, T. C., Pringouri, K., Stich, T. A., et al. (2012). Synthetic model of the asymmetric $[Mn_3CaO_4]$ cubane core of the oxygen-evolving complex of photosystem II. *Proc. Natl. Acad. Sci. U.S.A.* 109, 2257–2262.
- Noguchi, T. (2007). Light-induced FTIR difference spectroscopy as a powerful tool toward understanding the molecular mechanism of photosynthetic oxygen evolution. *Photosynth. Res.* 91, 59–69.
- Noguchi, T. (2008a). Fourier transform infrared analysis of the photosynthetic oxygen-evolving center. *Coord. Chem. Rev.* 252, 336–346.
- Noguchi, T. (2008b). FTIR detection of water reactions in the oxygen-evolving centre of photosystem II. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 363, 1189–1194.
- Noguchi, T., Ono, T., and Inoue, Y. (1992). Detection of structural changes upon S1-to-S2 transition in the oxygen-evolving manganese cluster in photosystem II by light-induced Fourier transform infrared difference spectroscopy. *Biochemistry* 31, 5953–5956.
- Noguchi, T., and Sugiura, M. (2000). Structure of an active water molecule in the water-oxidizing complex of photosystem II as studied by FTIR spectroscopy. *Biochemistry* 39, 10943–10949.
- Noguchi, T., and Sugiura, M. (2001). Flash-induced Fourier transform infrared detection of the structural changes during the S-state cycle of the oxygen-evolving complex in photosystem II. *Biochemistry* 40, 1497–1502.
- Noguchi, T., and Sugiura, M. (2002a). Flash-induced FTIR difference spectra of the water oxidizing complex in moderately hydrated photosystem II core films: effect of hydration extent on S-state transitions. *Biochemistry* 41, 2322–2330.
- Noguchi, T., and Sugiura, M. (2002b). FTIR detection of water reactions during the flash-induced S-state cycle of the photosynthetic water-oxidizing complex. *Biochemistry* 41, 15706–15712.
- Noguchi, T., Suzuki, H., Tsuno, M., Sugiura, M., and Kato, C. (2012). Time-resolved infrared detection of the proton and protein dynamics during photosynthetic oxygen evolution. *Biochemistry* 51, 3205–3214.
- Pecoraro, V., Baldwin, M., Caudle, M., Hsieh, W.-Y., and Law, N. (1998). A proposal for water oxidation in photosystem II. *Pure Appl. Chem.* 70, 925–930.
- Pushkar, Y., Yano, J., Sauer, K., Boussac, A., and Yachandra, V. K. (2008). Structural changes in the Mn_4Ca cluster and the mechanism of photosynthetic water splitting. *Proc. Natl. Acad. Sci. U.S.A.* 105, 1879–1884.
- Rapatskiy, L., Cox, N., Savitsky, A., Ames, W. M., Sander, J., Nowaczyk, M. M., et al. (2012). Detection of the water-binding sites of the oxygen-evolving complex of photosystem II using W-band ^{17}O electron-electron double resonance-detected NMR spectroscopy. *J. Am. Chem. Soc.* 134, 16619–16634.
- Service, R., Yano, J., McConnell, I., Hwang, H. J., Nicks, D., Hille, R., et al. (2010). Participation of glutamate-354 of the CP43 polypeptide in the ligation of Mn and the binding of substrate water in photosystem II. *Biochemistry* 50, 63–81.
- Shimada, Y., Suzuki, H., Tsuchiya, T., Mimuro, M., and Noguchi, T. (2011). Structural coupling of an arginine side chain with the oxygen-evolving Mn_4Ca cluster in photosystem II as revealed by isotope-edited Fourier transform infrared spectroscopy. *J. Am. Chem. Soc.* 133, 3808–3811.
- Shimada, Y., Suzuki, H., Tsuchiya, T., Tomo, T., Noguchi, T., and Mimuro, M. (2009). Effect of a single-amino acid substitution of the 43 kDa chlorophyll protein on the oxygen-evolving reaction of the cyanobacterium *Synechocystis* sp. PCC 6803: analysis of the Glu354Gln mutation. *Biochemistry* 48, 6095–6103.
- Siegbahn, P. E. M. (2009). Structures and energetics for O_2 formation in photosystem II. *Acc. Chem. Res.* 42, 1871–1880.
- Sproviero, E. M., Gascón, J. A., Mcevoy, J. P., Brudvig, G. W., and Batista, V. S. (2008). Computational studies of the O_2 -evolving complex of photosystem II and biomimetic oxomanganese complexes. *Coord. Chem. Rev.* 252, 395–415.
- Strickler, M. A., Hillier, W., and Debus, R. J. (2006). No evidence from FTIR difference spectroscopy that glutamate-189 of the D1 polypeptide ligates a Mn ion that undergoes oxidation during the S0 to S1, S1 to S2, or S2 to S3 transitions in photosystem II. *Biochemistry* 45, 8801–8811.
- Strickler, M. A., Hwang, H. J., Burnap, R. L., Yano, J., Walker, L. M., Service, R. J., et al. (2008). Glutamate-354 of the CP43 polypeptide interacts with the oxygen-evolving Mn_4Ca cluster of photosystem II: a preliminary characterization of the Glu354Gln mutant. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 363, 1179–1187.
- Strickler, M. A., Walker, L. M., Hillier, W., Britt, R. D., and Debus, R. J. (2007). No evidence from FTIR difference spectroscopy that aspartate-342 of the D1 polypeptide ligates a Mn ion that undergoes oxidation during the S0 to S1, S1 to S2, or S2 to S3 transitions in photosystem II. *Biochemistry* 46, 3151–3160.
- Strickler, M. A., Walker, L. M., Hillier, W., and Debus, R. J. (2005). Evidence from biosynthetically incorporated strontium and FTIR difference spectroscopy that the C-terminus of the D1 polypeptide of photosystem II does not ligate calcium. *Biochemistry* 44, 8571–8577.
- Suzuki, H., Sugiura, M., and Noguchi, T. (2009). Monitoring proton release during photosynthetic water oxidation in photosystem II by means of isotope-edited infrared spectroscopy. *J. Am. Chem. Soc.* 131, 7849–7857.
- Suzuki, H., Taguchi, Y., Sugiura, M., Boussac, A., and Noguchi, T. (2006). Structural perturbation of the carboxylate ligands to the manganese cluster upon Ca^{2+}/Sr^{2+} exchange in the S-state cycle of photosynthetic oxygen evolution as studied by flash-induced FTIR difference spectroscopy. *Biochemistry* 45, 13454–13464.
- Tsuno, M., Suzuki, H., Kondo, T., Mino, H., and Noguchi, T. (2011). Interaction and inhibitory effect of ammonium cation in the oxygen evolving center of photosystem II. *Biochemistry* 50, 2506–2514.
- Umena, Y., Kawakami, K., Shen, J. R., and Kamiya, N. (2011). Crystal structure of oxygen-evolving photosystem II at a resolution of 1.9 Å. *Nature* 473, 55–60.
- Visser, H., Dubé, C. E., Armstrong, W. H., Sauer, K., and Yachandra, V. K. (2002). FTIR spectra and normal-mode analysis of a tetranuclear manganese adamantane-like complex in two electrochemically prepared oxidation states: relevance to the oxygen-evolving complex of photosystem II. *J. Am. Chem. Soc.* 124, 11008–11017.
- Yamanari, T., Kimura, Y., Mizusawa, N., Ishii, A., and Ono, T.-A. (2004). Mid-to low-frequency Fourier transform infrared spectra of S-state cycle for photosynthetic water oxidation in *Synechocystis* sp. PCC 6803. *Biochemistry* 43, 7479–7490.
- Yano, J., Kern, J., Sauer, K., Latimer, M. J., Pushkar, Y., Biesiadka, J., et al. (2006). Where water is oxidized to dioxygen: structure of the photosynthetic Mn_4Ca cluster. *Science* 314, 821–825.

Conflict of Interest Statement: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 31 March 2013; paper pending published: 26 April 2013; accepted: 29 April 2013; published online: 21 May 2013.

Citation: Chu H-A (2013) Fourier transform infrared difference spectroscopy for studying the molecular mechanism of photosynthetic water oxidation. *Front. Plant Sci.* 4:146. doi: 10.3389/fpls.2013.00146

This article was submitted to *Frontiers in Plant Physiology*, a specialty of *Frontiers in Plant Science*.

Copyright © 2013 Chu. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in other forums, provided the original authors and source are credited and subject to any copyright notices concerning any third-party graphics etc.